

REVIFU

Expert Review of Precision Medicine and Drug **Development**

Personalized medicine in drug development and clinical practice

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/tepm20

The pharmacogenetics of tacrolimus and its implications for personalized therapy in kidney transplant recipients

Marith I. Francke, Brenda C.M. de Winter, Laure Elens, Nuria Lloberas & Dennis A. Hesselink

To cite this article: Marith I. Francke, Brenda C.M. de Winter, Laure Elens, Nuria Lloberas & Dennis A. Hesselink (2020) The pharmacogenetics of tacrolimus and its implications for personalized therapy in kidney transplant recipients, Expert Review of Precision Medicine and Drug Development, 5:5, 313-316, DOI: 10.1080/23808993.2020.1776107

To link to this article: <u>https://doi.org/10.1080/23808993.2020.1776107</u>



Published online: 19 Jun 2020.

ك

Submit your article to this journal

Article views: 1106



View related articles 🗹



View Crossmark data 🗹



Citing articles: 2 View citing articles 🕑

EDITORIAL

Check for updates

Taylor & Francis

Taylor & Francis Group

The pharmacogenetics of tacrolimus and its implications for personalized therapy in kidney transplant recipients

Marith I. Francke^{a,b}, Brenda C.M. de Winter^c, Laure Elens^d, Nuria Lloberas^e and Dennis A. Hesselink^{a,b}

^aDepartment of Internal Medicine, Division of Nephrology and Transplantation, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands; ^bRotterdam Transplant Group, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands; ^cDepartment of Hospital Pharmacy, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands; ^dLouvain Drug Research Institute, Université Catholique De Louvain, Louvain, Belgium; ^eDepartment of Nephrology, IDIBELL, Hospital Universitari Di Bellvitge, University of Barcelona, Barcelona, Spain

ARTICLE HISTORY Received 9 April 2020; Accepted 21 May 2020

KEYWORDS Tacrolimus; kidney transplantation; pharmacokinetics; pharmacogenetics; immunosuppression

Twenty-five years after its approval by the FDA and EMA, tacrolimus remains the cornerstone of immunosuppressive treatment following solid organ transplantation [1,2]. The drug is highly effective in preventing acute rejection but its clinical use still is complicated by its narrow therapeutic range and high inter- and intra-patient pharmacokinetic variability [3,4]. In many transplant centers across the world, the starting dose of tacrolimus is based on a patient's bodyweight, followed by whole-blood tacrolimus concentration measurement (preferably when the drug is in steady state) and dose titration. Therapeutic drug monitoring (TDM) is considered standard of care and limits the time that a patient is under- or overexposed to tacrolimus [1]. However, TDM is a reactive approach and in clinical practice, dosing of tacrolimus remains trial and error. Bodyweight is a poor predictor of an individual's tacrolimus clearance [5,6] and as a result, it may take up to three weeks before a patient has a tacrolimus exposure within the target concentration range [7,8]. In addition, an individual's tacrolimus metabolic phenotype has been associated with transplantation outcomes [9]. A low tacrolimus concentration-to-dose ratio (C/D ratio), which indicates a more rapid tacrolimus clearance, is an independent risk factor for death-censored graft survival [9]. There is an unmet need to personalize tacrolimus treatment and limit the time that transplant recipients are under- or overexposed to tacrolimus in order to limit the risks of acute rejection and tacrolimus-related toxicity, respectively [10].

The inter-individual differences in tacrolimus pharmacokinetics are in large part determined genetically. Single-nucleotide polymorphisms (SNPs) in multiple genes have been associated with tacrolimus dose requirement (for a review see references [11,12]). Especially, SNPs in the cytochrome P450 (CYP) *3A4* and *CYP3A5* genes, which encode the main tacrolimus-metabolizing enzymes, have been strongly linked to an individual's drug metabolizing phenotype [6,13,14]. Carriers of at least one *CYP3A5*1* allele are considered CYP3A5 expressers and require a tacrolimus dose that is at least 1.5 times higher compared to so-called CYP3A5 non-expressers (individuals with the *CYP3A5*3/*3* genotype) [1]. The *CYP3A4*22* SNP has been associated with slower tacrolimus

clearance and a lower dose requirement [14–18]. In addition, the *CYP3A4*20* SNP [16,19] and genetic variations in the *ABCB1, POR, PPARA* and NR112 genes have all been associated with tacrolimus pharmacokinetics. The effect of these variants seems smaller than that of *CYP3A4* and *CYP3A5* and is probably less clinically relevant [20,21].

It was hoped that pharmacogenetics-guided tacrolimus dosing would improve clinical tacrolimus treatment [22]. In particular, a pharmacogenetics-based approach may lead to a more appropriate tacrolimus starting dose, thereby limiting the time a patient is outside the target concentration range in the critical early post-transplant phase [23]. The genotype-based tacrolimus efficacy of dosing was investigated in three randomized-controlled clinical trials [7,8,24]. In all three trials, the tacrolimus starting dose was based on an individual transplant recipient's CYP3A5 genotype, being higher in CYP3A5 expressers (carriers of a *1 allele) and lower in non-expressers. In the study by Thervet et al. [8] genotype-guided tacrolimus dosing resulted in significantly more patients being on target at first-steady state (i.e. after 5 unaltered tacrolimus doses) compared with the control group who received a standard, bodyweightbased tacrolimus starting dose (43.2% versus 29.1%; p = 0.03). In the study by Min et al. [24] this approach significantly shortened the time to reach the tacrolimus target concentration range in pediatric solid organ transplant recipients (n = 53 patients). However, the absolute difference was small (a median of 3.4 versus 4.7 days in the genotype-guided versus the standard dose group, respectively). In the trial by Shuker et al. [7] there was no difference in the proportion of living donor kidney transplant recipients that were on target at first steady state measurement. In the standard, bodyweight-based dose tacrolimus group, 37.4% of patients was on target (10--15 ng/mL) at day three after transplantation, whereas this was 35.6% in the experimental arm in which the tacrolimus starting dose was based on CYP3A5 genotype. Importantly,

CONTACT Marith I. Francke Improve m.francke@erasmusmc.nl Department of Internal Medicine, Division of Nephrology and Transplantation, Erasmus MC, University Medical Center Rotterdam, The Netherlands

 $[\]ensuremath{\mathbb{C}}$ 2020 Informa UK Limited, trading as Taylor & Francis Group

in none of these three trials any clinical benefit of *CYP3A5*guided tacrolimus dosing was observed, either in the shortor long-term [7,8,24,25]. The lack of evidence for clinical benefit of genotype-based tacrolimus dosing, is one of the reasons why this strategy has not been widely implemented clinically [26]. The Clinical Pharmacogenetics Implementation Consortium recommends to only use pharmacogenetics-based tacrolimus dosing when an individual's genotype is already available [27].

Not only a patient's genotype but also other factors affect tacrolimus pharmacokinetics, including age, plasma albumin concentration, body surface area, co-medication, ethnicity, and hematocrit [10]. It is reasonable to assume that combining these factors with genetic information in a population pharmacokinetic (popPK) model will allow clinicians to predict a patient's tacrolimus dose requirement with more precision than a prediction based on genotype (or bodyweight) alone, which might increase the clinical benefit. To date, multiple models predicting the tacrolimus starting dose have been developed, both for pediatric, as well as for adult renal transplant recipients [3,28-31]. However, few of these popPK models were externally validated [3,28,29,32,33] and even fewer were tested prospectively [34,35]. The latter is essential before model-based tacrolimus dosing can be implemented clinically. We developed a popPK model for pediatric kidney transplant recipients and validated this model both internally and externally [28]. However, when tested prospectively, the model did not predict a patient's tacrolimus dose requirement sufficiently [34]. Currently, several prospective intervention trials, investigating the efficacy of initial algorithm-based dosing in adult renal allograft recipients are ongoing (https://www.trialregister.nl/trial/7360; NCT03465410; NCT03020589; NCT03527238 see clinicaltrials.gov).

Recent research has mainly focused on the calculation of the tacrolimus starting dose. However, not only the achievement but also the maintenance of the tacrolimus exposure within the target concentration range is a challenge. Patients with a tacrolimus concentration within the therapeutic range at first steady state can have a tacrolimus concentration that lies outside this range on subsequent days. This intra-patient variability is in part explained by post-transplant changes in hematocrit (e.g. resulting from blood loss due to surgery or frequent sampling) and albumin concentration (due to, for example, recovery of liver function and co-existing inflammation), and hepatic metabolism (from loss of uremic toxins [36]). Computerized follow-up dosing based on popPK models, may better maintain the targeted tacrolimus exposure and further individualize tacrolimus treatment after transplantation. Størset et al. [37] reported a clinical trial in which standard or high-immunological risk kidney transplant recipients were randomized (after initial bodyweight-based dosing) to undergo either computerized follow-up tacrolimus dosing or conventional tacrolimus dosing by experienced transplant physicians. Computerized dosing, based on several variables (historical tacrolimus dosages, previously measured tacrolimus concentrations, fat-free mass, hematocrit, and time after transplantation), significantly improved the achievement of tacrolimus concentrations within the target range. During the first eight weeks after transplantation, a median of 90% of the patients' tacrolimus concentrations was within the target range following computerized dosing, whereas this was 78% following standard dosing in the standard risk group (p < 0.001). Also, in the high-risk group, a significantly higher proportion of the patients' tacrolimus concentrations was within the target range following computerized dosing compared to standard dosing (median 77% *versus* 59%, respectively; p = 0.04). In the computer-based dosing group, a lower two-hour plasma glucose concentration was observed following an oral glucose tolerance test, performed eight weeks post-transplantation. This indicates a possible beneficial clinical effect of algorithm-based tacrolimus dosing and the resulting reduced supra-therapeutic exposure in terms of glucose metabolism [37].

Another strategy which may optimize tacrolimus treatment and increase clinical benefit is the monitoring of tacrolimus in a matrix other than whole-blood. Contradictory results have been reported on the correlation between whole-blood tacrolimus exposure and both rejection and drug-related toxicity [38-41]. The tacrolimus concentration at its target site (within lymphocytes) may better correlate with efficacy and clinical outcomes. This hypothesis was supported by the study of Capron et al., in which the tacrolimus concentration within peripheral-blood mononuclear cells correlated significantly with the occurrence and severity of rejection in liver transplant recipients [42]. These findings need to be substantiated in larger studies. Interestingly, the intracellular tacrolimus concentration has been related, although not consistently, to genetic variability in CYP3A4 and CYP3A5 but also to variability in the expression of drug transporter proteins such as ABCB1, which determine the drug's distribution [43-45].

Expert opinion

Although the application of popPK models has the potential to move the field forward, it is clear that current models, which include demographic and genetic patient characteristics, do not explain all inter- and intra-patient variability in tacrolimus pharmacokinetics. Recently, Zimmermann et al. [46] showed that residual variability in drug metabolism may be explained by variation in the human microbiome. Microbiome-encoded enzymes contribute substantially to pre-systemic metabolism of many drugs [46], including tacrolimus [47,48]. Lee et al. [47] demonstrated that the abundance of fecal Faecalibacterium prausnitzii is associated with higher tacrolimus requirement in the early post-transplantation phase. We feel that a further exploration of the role of the human microbiome in tacrolimus pharmacokinetics is an important research topic for the years to come and has the potential to improve the performance of future popPK models. Such studies are at present underway (see clinical trials.gov, NCT04207177). Hopefully, the revolutionary findings of Zimmerman et al. and Lee et al. [46,47] will fuel the transplant community to optimize tacrolimus treatment further. We owe this to our patients because tacrolimus will likely remain an essential component of immunosuppressive therapy after transplantation in the decade to come.

Funding

This paper was not funded.

Declaration of interest

D.A. Hesselink has received grant support (paid to his institution) from Astellas Pharma, Chiesi Farmaceutici SpA and Bristol Myers-Squibb, as well as lecture and consulting fees from Astellas Pharma, Chiesi Farmaceutici SpA, Novartis Pharma and Vifor Pharma. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

Reviewers disclosure

Peer reviewers on this manuscript have no relevant financial relationships or otherwise to disclose.

References

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- Brunet M, van Gelder T, Asberg A, et al. Therapeutic drug monitoring of tacrolimus-personalized therapy: second consensus report. Ther Drug Monit. 2019;41:261–307.
- Up-to-date consensus paper on the use and measurement of tacrolimus.
- Starzl TE, Todo S, Fung J, et al. FK 506 for liver, kidney, and pancreas transplantation. Lancet. 1989;2:1000–1004.
- Andrews LM, Hesselink DA, van Schaik RHN, et al. A population pharmacokinetic model to predict the individual starting dose of tacrolimus in adult renal transplant recipients. Br J Clin Pharmacol. 2019;85:601–615.
- 4. Shuker N, van Gelder T, Hesselink DA. Intra-patient variability in tacrolimus exposure: causes, consequences for clinical management. Transplant Rev (Orlando). 2015;29:78–84.
- Andrews LM, de Winter BC, Tang JT, et al. Overweight kidney transplant recipients are at risk of being overdosed following standard bodyweight-based tacrolimus starting dose. Transplant Direct. 2017;3:e129.
- Press RR, Ploeger BA, den Hartigh J, et al. Explaining variability in tacrolimus pharmacokinetics to optimize early exposure in adult kidney transplant recipients. Ther Drug Monit. 2009;31:187–197.
- Shuker N, Bouamar R, van Schaik RH, et al. A randomized controlled trial comparing the efficacy of Cyp3a5 genotype-based with body-weight-based tacrolimus dosing after living donor kidney transplantation. Am J Transplant. 2016;16:2085–2096.
- Thervet E, Loriot MA, Barbier S, et al. Optimization of initial tacrolimus dose using pharmacogenetic testing. Clin Pharmacol Ther. 2010;87:721–726.
- 9. Jouve T, Fonrose X, Noble J, et al. The TOMATO study (TacrOlimus MetabolizAtion in kidney TransplantatiOn): impact of the concentration-dose ratio on death-censored graft survival. Transplantation. 2020;104(6):1263–1271.
- Large clinical study demonstrating a correlation between tacrolimus C/D ratio and graft survival.
- Staatz CE, Tett SE. Clinical pharmacokinetics and pharmacodynamics of tacrolimus in solid organ transplantation. Clin Pharmacokinet. 2004;43:623–653.
- 11. Hesselink DA, Bouamar R, Elens L, et al. The role of pharmacogenetics in the disposition of and response to tacrolimus in solid organ transplantation. Clin Pharmacokinet. 2014;53:123–139.
- Picard N, Bergan S, Marquet P, et al. Pharmacogenetic biomarkers predictive of the pharmacokinetics and pharmacodynamics of immunosuppressive drugs. Ther Drug Monit. 2016;38(Suppl 1):S57–69.
- 13. Hesselink DA, van Schaik RH, van der Heiden IP, et al. Genetic polymorphisms of the CYP3A4, CYP3A5, and MDR-1 genes and pharmacokinetics of the calcineurin inhibitors cyclosporine and tacrolimus. Clin Pharmacol Ther. 2003;74:245–254.

- 14. Lloberas N, Elens L, Llaudo I, et al. The combination of CYP3A4*22 and CYP3A5*3 single-nucleotide polymorphisms determines tacrolimus dose requirement after kidney transplantation. Pharmacogenet Genomics. 2017;27:313–322.
- 15. Elens L, Capron A, van Schaik RH, et al. Impact of CYP3A4*22 allele on tacrolimus pharmacokinetics in early period after renal transplantation: toward updated genotype-based dosage guidelines. Ther Drug Monit. 2013;35:608–616.
- Gomez-Bravo MA, Apellaniz-Ruiz M, Salcedo M, et al. Influence of donor liver CYP3A4*20 loss-of-function genotype on tacrolimus pharmacokinetics in transplanted patients. Pharmacogenet Genomics. 2018;28:41–48.
- Elens L, Haufroid V. Genotype-based tacrolimus dosing guidelines: with or without CYP3A4*22? Pharmacogenomics. 2017;18:1473–1480.
- Woillard JB, Mourad M, Neely M, et al. Tacrolimus updated guidelines through popPK modeling: how to benefit more from CYP3A pre-emptive genotyping prior to kidney transplantation. Front Pharmacol. 2017;8:358.
- Lloberas N, Hesselink DA, van Schaik RH, et al. Detection of a rare CYP3A4 variant in a transplant patient characterized by a tacrolimus poor metabolizer phenotype. Pharmacogenomics. 2018;19:305–310.
- Lunde I, Bremer S, Midtvedt K, et al. The influence of CYP3A, PPARA, and POR genetic variants on the pharmacokinetics of tacrolimus and cyclosporine in renal transplant recipients. Eur J Clin Pharmacol. 2014;70:685–693.
- 21. Li JL, Liu S, Fu Q, et al. Interactive effects of CYP3A4, CYP3A5, MDR1 and NR112 polymorphisms on tracrolimus trough concentrations in early postrenal transplant recipients. Pharmacogenomics. 2015;16:1355–1365.
- 22. van Gelder T, Hesselink DA. Dosing tacrolimus based on CYP3A5 genotype: will it improve clinical outcome? Clin Pharmacol Ther. 2010;87:640–641.
- MacPhee IA, Fredericks S, Tai T, et al. The influence of pharmacogenetics on the time to achieve target tacrolimus concentrations after kidney transplantation. Am J Transplant. 2004;4:914–919.
- 24. Min S, Papaz T, Lafreniere-Roula M, et al. A randomized clinical trial of age and genotype-guided tacrolimus dosing after pediatric solid organ transplantation. Pediatr Transplant. 2018;22: e13285.
- Pallet N, Etienne I, Buchler M, et al. Long-term clinical impact of adaptation of initial tacrolimus dosing to CYP3A5 genotype. Am J Transplant. 2016;16:2670–2675.
- 26. Koch BC, van Schaik RH, van Gelder T, et al. Doubt about the feasibility of preemptive genotyping. Clin Pharmacol Ther. 2013;93:233.
- Birdwell KA, Decker B, Barbarino JM, et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines for CYP3A5 genotype and tacrolimus dosing. Clin Pharmacol Ther. 2015;98:19–24.
- Andrews LM, Hesselink DA, van Gelder T, et al. A population pharmacokinetic model to predict the individual starting dose of tacrolimus following pediatric renal transplantation. Clin Pharmacokinet. 2018;57:475–489.
- Andreu F, Colom H, Elens L, et al. A new CYP3A5*3 and CYP3A4*22 cluster influencing tacrolimus target concentrations: a population approach. Clin Pharmacokinet. 2017;56:963–975.
- Andrews LM, Riva N, de Winter BC, et al. Dosing algorithms for initiation of immunosuppressive drugs in solid organ transplant recipients. Expert Opin Drug Metab Toxicol. 2015;11:921–936.
- 31. Han N, Yun HY, Hong JY, et al. Prediction of the tacrolimus population pharmacokinetic parameters according to CYP3A5 genotype and clinical factors using NONMEM in adult kidney transplant recipients. Eur J Clin Pharmacol. 2013;69:53–63.
- Boughton O, Borgulya G, Cecconi M, et al. A published pharmacogenetic algorithm was poorly predictive of tacrolimus clearance in an independent cohort of renal transplant recipients. Br J Clin Pharmacol. 2013;76:425–431.
- Passey C, Birnbaum AK, Brundage RC, et al. Dosing equation for tacrolimus using genetic variants and clinical factors. Br J Clin Pharmacol. 2011;72:948–957.

- 34. Andrews LM, de Winter BCM, Cornelissen EAM, et al. A population pharmacokinetic model does not predict the optimal starting dose of tacrolimus in pediatric renal transplant recipients in a prospective study: lessons learned and model improvement. Clin Pharmacokinet. 2020;59(5):591–603.
- 35. Chen SY, Li JL, Meng FH, et al. Individualization of tacrolimus dosage basing on cytochrome P450 3A5 polymorphism–a prospective, randomized, controlled study. Clin Transplant. 2013;27:E272–281.
- Yeung CK, Shen DD, Thummel KE, et al. Effects of chronic kidney disease and uremia on hepatic drug metabolism and transport. Kidney Int. 2014;85:522–528.
- 37. Storset E, Asberg A, Skauby M, et al. Improved tacrolimus target concentration achievement using computerized dosing in renal transplant recipients-a prospective, randomized study. Transplantation. 2015;99:2158–2166.
- Bouamar R, Shuker N, Hesselink DA, et al. Tacrolimus predose concentrations do not predict the risk of acute rejection after renal transplantation: a pooled analysis from three randomized-controlled clinical trials(dagger). Am J Transplant. 2013;13:1253–1261.
- Randomized controlled clinical trial demonstrating the superiority of computer over human-based tacrolimus dosing.
- Rodriguez-Peralvarez M, Germani G, Darius T, et al. Tacrolimus trough levels, rejection and renal impairment in liver transplantation: a systematic review and meta-analysis. Am J Transplant. 2012;12:2797–2814.
- 40. Gaynor JJ, Ciancio G, Guerra G, et al. Lower tacrolimus trough levels are associated with subsequently higher acute rejection risk during the first 12 months after kidney transplantation. Transpl Int. 2016;29:216–226.
- 41. Israni AK, Riad SM, Leduc R, et al. Tacrolimus trough levels after month 3 as a predictor of acute rejection following kidney

transplantation: a lesson learned from DeKAF genomics. Transpl Int. 2013;26:982–989.

- 42. Capron A, Lerut J, Latinne D, et al. Correlation of tacrolimus levels in peripheral blood mononuclear cells with histological staging of rejection after liver transplantation: preliminary results of a prospective study. Transpl Int. 2012;25:41–47.
- First study demonstrating a correlation between clinical outcomes and intracellular tacrolimus concentration in solid organ transplantation.
- Tron C, Woillard JB, Houssel-Debry P, et al. Pharmacogenetic-Whole blood and intracellular pharmacokinetic-Pharmacodynamic (PG-PK2-PD) relationship of tacrolimus in liver transplant recipients. PLoS One. 2020;15:e0230195.
- 44. Capron A, Mourad M, De Meyer M, et al. CYP3A5 and ABCB1 polymorphisms influence tacrolimus concentrations in peripheral blood mononuclear cells after renal transplantation. Pharmacogenomics. 2010;11:703–714.
- 45. Han SS, Yang SH, Kim MC, et al. Monitoring the intracellular tacrolimus concentration in kidney transplant recipients with stable graft function. PLoS One. 2016;11:e0153491.
- 46. Zimmermann M, Zimmermann-Kogadeeva M, Wegmann R, et al. Mapping human microbiome drug metabolism by gut bacteria and their genes. Nature. 2019;570:462–467.
- •• Study demonstrating the importance of the human microbiome for drug metabolism.
- Lee JR, Muthukumar T, Dadhania D, et al. Gut microbiota and tacrolimus dosing in kidney transplantation. PLoS One. 2015;10:e0122399.
- •• Study on the relationship between the human microbiome and tacrolimus dosing.
- Guo Y, Crnkovic CM, Won KJ, et al. Commensal gut bacteria convert the immunosuppressant tacrolimus to less potent metabolites. Drug Metab Dispos. 2019;47:194–202.