








Genetic Variation in *HSD17B13* Reduces the Risk of Developing Cirrhosis and Hepatocellular Carcinoma in Alcohol Misusers

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BACKGROUND AND AIMS: Carriage of rs738409:G in *patatin-like phospholipase domain containing 3* (*PNPLA3*) is associated with an increased risk for developing alcohol-related cirrhosis and hepatocellular carcinoma (HCC). Recently, rs72613567:TA in *hydroxysteroid 17-beta dehydrogenase 13* (*HSD17B13*) was shown to be associated with a reduced risk for developing alcohol-related liver disease and to attenuate the risk associated with carriage of *PNPLA3* rs738409:G. This study explores the risk associations between these two genetic variants and the development of alcohol-related cirrhosis and HCC.

APPROACH AND RESULTS: Variants in *HSD17B13* and *PNPLA3* were genotyped in 6,171 participants, including 1,031 with alcohol-related cirrhosis and HCC, 1,653 with alcohol-related cirrhosis without HCC, 2,588 alcohol misusers with no liver disease, and 899 healthy controls. Genetic associations with the risks for developing alcohol-related cirrhosis and HCC were determined using logistic regression analysis. Carriage of *HSD17B13* rs72613567:TA was associated with a lower risk for developing both cirrhosis (odds ratio [OR], 0.79; 95% confidence interval [CI], 0.72-0.88; $P = 8.13 \times 10^{-6}$) and HCC (OR, 0.77; 95% CI, 0.68-0.89;

$P = 2.27 \times 10^{-4}$), whereas carriage of *PNPLA3* rs738409:G was associated with an increased risk for developing cirrhosis (OR, 1.70; 95% CI, 1.54-1.88; $P = 1.52 \times 10^{-26}$) and HCC (OR, 1.77; 95% CI, 1.58-1.98; $P = 2.31 \times 10^{-23}$). These associations remained significant after adjusting for age, sex, body mass index, type 2 diabetes, and country. Carriage of *HSD17B13* rs72613567:TA attenuated the risk for developing cirrhosis associated with *PNPLA3* rs738409:G in both men and women, but the protective effect against the subsequent development of HCC was only observed in men (OR_{allelic}, 0.75; 95% CI, 0.64-0.87; $P = 1.72 \times 10^{-4}$).

CONCLUSIONS: Carriage of variants in *PNPLA3* and *HSD17B13* differentially affect the risk for developing advanced alcohol-related liver disease. A genotypic/phenotypic risk score might facilitate earlier diagnosis of HCC in this population. (HEPATOLOGY 2020;0:1-15).

Alcohol-related liver disease has a global prevalence of 12% and accounts for half of all cirrhosis-associated deaths in Europe and the

Abbreviations: BMI, body mass index; CI, confidence interval; HBV, hepatitis B virus; HCC, hepatocellular carcinoma; HCV, hepatitis C virus; *HSD17B13*, hydroxysteroid 17-beta dehydrogenase 13; MAF, minor allele frequency; NAFLD, nonalcoholic fatty liver disease; OR, odds ratio; PAF, population-attributable fraction; *PNPLA3*, patatin-like phospholipase domain containing 3; SNP, single-nucleotide polymorphism; T2DM, type 2 diabetes; TM6SF2, transmembrane 6 superfamily member 2.

Received August 19, 2019; accepted September 18, 2019.

Additional Supporting Information may be found at onlinelibrary.wiley.com/doi/10.1002/hep.30996/supinfo.

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United States.⁽¹⁻³⁾ Alcohol-related cirrhosis is now the second-most-common indication for liver transplantation, accounting for approximately 40% of all primary liver transplants in Europe and approximately 25% in the United States.⁽⁴⁾

Chronic alcohol misuse is associated with the development of a broad spectrum of liver injury. Hepatic steatosis develops in most heavy alcohol users, but more substantial liver injury only develops with persistent alcohol misuse over time; inflammation and progressive

fibrosis will develop in 10%-35% of individuals, whereas cirrhosis is observed in only 10%-15%.⁽⁵⁻⁷⁾ Between 5% and 15% of people with alcohol-related cirrhosis are at risk for developing hepatocellular carcinoma (HCC); the annual incidence is 2.5%-3.0%, with a 5-year cumulative risk of around 8%.⁽⁸⁻¹¹⁾ The global incidence of HCC is increasing; it is now the fifth-most-frequent cancer and third-most-frequent cause of cancer-related mortality worldwide; one third of cases develop on a background of alcohol-related cirrhosis.⁽¹²⁾

Supported by grants from the Swiss National Funds (SNF no. 310030_169196) and the Swiss Foundation for Alcohol Research (SSA) to F. S., and the Liver Systems Medicine (LiSyM) Network funded by the German Federal Ministry for Education and Research (BmBF) to J. H. H. D. N. and U. S. were supported by a grant from the Deutsche Krebshilfe (70112169).

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View this article online at wileyonlinelibrary.com.

DOI 10.1002/hep.30996

Potential conflict of interest: Dr. Weiss advises and is on the speakers' bureau for Chiesi. He consults for Univar and Vivet. He advises GMPO, is on the speakers' bureau for AbbVie and Novartis, and received grants from Astellas. Dr. Wege consults for Bayer, Bristol-Myers Squibb, Lilly, Eisai, MSD, and Ipsen. Dr. Berg advises, is on the speakers' bureau for, and received grants from AbbVie, Alexion, Bayer, Gilead, Eisai, Intercept, Ipsen, Janssen, MSD/Merck, Novartis, and Sequana.

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The susceptibility to develop significant alcohol-related liver disease is determined by the interplay of a number of risk factors, including sex; ethnicity; the amount/pattern of alcohol drinking; coffee consumption; cigarette smoking; comorbidities such as obesity, type 2 diabetes, and hepatitis C virus (HCV) infection; and a number of host genetic factors.⁽¹³⁾ Carriage of the common missense variant rs738409:G in *patatin-like phospholipase domain containing 3* (*PNPLA3*) is the most robustly validated risk locus for the development of alcohol-related cirrhosis and HCC,⁽¹⁴⁻¹⁹⁾ accounting for 26.6%⁽¹⁴⁾ and 43.5%⁽¹⁸⁾ of the variance, respectively. Two further gene variants, rs58542926 in *transmembrane 6 superfamily member 2* (*TM6SF2*) and rs641738 in *membrane-bound O-acetyltransferase domain containing 7* (*MBOAT7*), are additional risk factors, albeit with much lower effect sizes.⁽¹⁹⁾

Recently, Abul-Husn et al.,⁽²⁰⁾ identified a splice variant rs72613567 in *hydroxysteroid 17-beta dehydrogenase 13* (*HSD17B13*) that appears to protect against the development of chronic liver injury in people for European ancestry. *HSD17B13* encodes an hepatic lipid droplet protein; the insertion of an adenine adjacent to the donor splice site on exon 6 (rs72613567:TA allele) results in an unstable and truncated protein with reduced enzymatic activity, suggesting a loss-of-function variant.⁽²⁰⁾ Abul-Husn et al.⁽²⁰⁾ found that carriage of *HSD17B13* rs72613567:TA was associated with reduced serum aminotransferase activities and a reduced risk for developing alcohol-related and nonalcoholic fatty liver disease (NAFLD) and, more specifically, for developing both alcohol-related and NAFLD-related cirrhosis.⁽²⁰⁾ The association with alcohol-related cirrhosis was the most compelling, but the total number of cases was very small. This group also showed that carriage of this variant attenuated the risk for developing progressive liver injury associated with carriage of rs738409:G in *PNPLA3*.⁽²⁰⁾

Yang et al.⁽²¹⁾ recently confirmed the association between carriage of rs72613567:TA in *HSD17B13* and a reduction in the risk for developing alcohol-, NAFLD-, and HCV-related liver disease and, more specifically, the risk for developing cirrhosis. They also found that carriage of this variant protected against the development of HCC, but only in patients with a background of alcohol-related liver disease. The sample size was small, and those with HCC were more frequently male and older than their counterparts without HCC. Thus, in order to validate these

findings, much larger cohorts are needed to allow for adjustment for these potential confounders.⁽²¹⁾ Interactions between variants in *HSD17B13* and *PNPLA3* were not addressed in this study.

The aims of the present study were to determine whether: (i) carriage of rs72613567:TA in *HSD17B13* protects against the development of alcohol-related cirrhosis and alcohol-related HCC; and, (ii) to explore possible risk interactions between rs72613567:TA in *HSD17B13* and rs738409:G in *PNPLA3*.

Patients and Methods

STUDY COHORTS

The study population consisted of 6,171 participants drawn from 1 of 4 study cohorts:

1. Alcohol-related cirrhosis and HCC (HCC; n = 1,031)
2. Alcohol-related cirrhosis without HCC (CIRR; n = 1,653)
3. Alcohol misusers with no evident liver disease (ALC; n = 2,588)
4. Healthy controls with no history of alcohol misuse or liver disease (n = 899)

Participants in cohorts 1-3 were recruited from hepatology units and addiction centers across Europe and were of self-reported Swiss/German/Austrian/Italian/British ancestry. Ninety of the UK samples with cirrhosis and HCC were obtained from the Nottingham Digestive Diseases Centre Biomedical Research Unit Research Tissue Bank (Rec Ref: 14/WA/1234). Participants in cohort 4 were recruited from the United Kingdom.

The diagnosis of alcohol-related cirrhosis was established as described in detail previously;⁽¹⁹⁾ briefly, the diagnosis was based on a history of prolonged, sustained alcohol intake of a minimum of 40 g/day in women and 60 g/day in men together with histological examination of liver tissue or compatible historical, clinical, laboratory, radiological, and endoscopic features of advanced chronic liver disease. Patients were excluded if they had any other potential cause of liver injury, specifically, if they were positive for hepatitis B surface antigen, anti-HCV immunoglobulin G, antinuclear antibodies (titer > 1:80), or antimitochondrial antibodies

(titer > 1:40). Patients with elevated serum ferritin concentrations and a transferrin saturation of >50%, a serum caeruloplasmin of <20 mg/dL (0.2 g/dL), or a serum alpha-1 antitrypsin of <70 mg/dL (13 μ mol/L) were further investigated and excluded, as appropriate.

The diagnosis of HCC was based on histological examination of tumor tissue or evidence on imaging, preferably using two modalities, of lesions that were hypervascular in the arterial phase with washout in the portal venous or delayed phases.⁽²²⁾ The severity of the underlying cirrhosis was assessed using Pugh's modification of Child's grading system.⁽²³⁾

The patients with alcohol misuse but no evidence of significant liver injury were recruited as described in detail previously;⁽¹⁹⁾ in brief, they had a background of alcohol consumption of at least 60 g/day for women and 80 g/day for men for ≥ 10 years with or without features of alcohol dependence.⁽²⁴⁾ None had historical, clinical, or laboratory evidence of liver disease, and its absence was confirmed either by a liver stiffness measurement (FibroScan, Echosens, Paris) of below 6 kPa (interquartile range <20%) or by the absence of histological liver damage.

Healthy controls were recruited from London branches of the National Health Service blood transfusion service, from general practitioners' surgeries, from among university students, and from the general public. None currently drank alcohol above a weekly maximum of 112 g for women and 168 g for men, nor had they done so at any time in the past. None had a history or clinical evidence of liver disease.

DNA PREPARATION AND GENOTYPING

Genomic DNA was extracted from venous blood samples and quantified using standard procedures.⁽¹⁹⁾ Genotyping of the single-nucleotide polymorphisms (SNP) of interest *viz.* *PNPLA3* rs738409 (Assay ID: C_7241_10) and *HSD17B13* rs72613567 (primer and probe sets manufactured through custom TaqMan Assay design) was performed using TaqMan SNP genotyping assays and chemistries (Applied Biosystems, Waltham, MA) on an automated platform with Tecan Freedom EVO and 384-well TeMO liquid-handling robots (Tecan, Männedorf, Switzerland) as described previously.^(18,19)

All process data were logged and administered with a database-driven LIMS. Reactions were completed and read in a 7900 HT TaqMan sequence detector system (Applied Biosystems, Waltham, MA). The amplification reaction was carried out with the TaqMan universal master mix at cycling conditions of 1 cycle for 10 minutes at 95°C, followed by 45 cycles for 15 seconds at 95°C and 1 minute at 60°C.

STATISTICAL ANALYSIS

Logistic regression and SNP*SNP interaction analyses were performed using SPSS v.25.0 (IBM Corporation, Armonk, NY). A three-way case control design was adopted: HCC versus CIRR; CIRR versus ALC; HCC versus ALC. Genotypic and allelic tests of association were assessed using two logistic regression models:

Model A: univariate logistic regression, and

Model B: adjustments for age, sex, body mass index (BMI), type 2 diabetes, and country.

Results, expressed as odds ratios (ORs) with their 95% confidence intervals (CI), were derived from beta coefficients and their standard deviations. Nominal two-sided asymptotic *P* values are reported for all tests.

The interactions between *HSD17B13* and *PNPLA3* were examined by logistic regression for the univariate additive and genotypic regression models, including both main SNP effects and the SNP*SNP interaction term.

Fixed-effect model meta-analysis using the inverse variance-weighted method to summarize effect sizes and forest plots were performed using the R package "metaphor" v.2.0-0.

Sex-specific *post hoc* analyses for the risk associations with *HSD17B13* and *PNPLA3* were performed. The Mantel-Haenszel test for trend was applied for testing a linear trend in observed genotype proportions from contingency tables for ALC < CIRR < HCC, by sex.

The population-attributable fraction (PAF) provides an epidemiological estimate of the proportion of a disorder that is attributable to a given risk factor. Thus, in this instance, it provides an estimate of how much lower the frequency of HCC would be in patients with alcohol-related cirrhosis if the risk genotype(s) were eliminated from the population.

The PAF was estimated for heterozygous and homozygous carriage using the formula²⁵:

$$(x-1)/x$$

where $x = (1 - p)^2 + 2p(1 - p)OR1 + p^2OR2^{(19)}$; p is the allele frequency in the CIRR or ALC cohorts, and OR1 and OR2 are the ORs associated with hetero- and homozygosity.

Combined PAF estimates were calculated as $PAF = 1 - (1 - PAF1)(1 - PAF2)(1 - PAFn)$ based on the individual PAFs for each associated SNP, assuming no multiplicative interaction between them.

ETHICS

The study protocol was approved by the ethics committees of the participating institutions; all included subjects provided written informed consent before inclusion into the study.

Results

The patient cohorts were predominantly male and middle aged. The patients with cirrhosis were

generally older than those misusing alcohol and more likely to be overweight and to have type 2 diabetes. The patients with cirrhosis and HCC were generally older than the patients with cirrhosis without malignant transformation and were proportionately more likely again to be male, overweight, and to have type 2 diabetes (Table 1). Laboratory variables showed the expected gradients (Supporting Table S1).

Genotyping was completed for both SNPs with call rates greater than 95% (Supporting Table S2). All markers followed Hardy-Weinberg equilibrium (cut-off HWE $P > 0.05$).⁽²⁶⁾ In the healthy controls, the minor allele frequencies (MAFs) for both *HSD17B13* rs72613567:TA (0.276) and *PNPLA3* rs738409 G (0.224) were comparable with those in public databases⁽²⁷⁾ and previous publications⁽¹⁸⁻²⁰⁾ and did not differ significantly from the MAFs in the ALC group (Supporting Table S2).

A progressive reduction in MAFs for *HSD17B13* rs72613567:TA was observed between the 3 groups: ALC (26.4%); CIRR (22.0%), and HCC (17.7%; $P_{\text{trend}} = 1.09 \times 10^{-15}$; Supporting Table S2). This contrasted with the expected stepwise increase in the MAFs for *PNPLA3* rs738409:G in the same 3 groups,

TABLE 1. Demographic Characteristics of the Study Cohorts

Cohorts	Number	Age [SD]	Men (%)	BMI [SD]	T2DM (%)	Nicotine (% Users)
Alcohol-related cirrhosis with HCC (HCC)						
Total	1,031	62 [10]*	91*	27.8 [4.8]*	45%*	47%*
Germany	778	61 [10]	91	27.8 [4.8]	45%	50%
Switzerland	115	61 [11]	85	27.9 [5.0]	53%	38%
United Kingdom [‡]	65	65 [9]	93	25.6 [3.1]	0%	N/A
Italy	73	72 [8]	92	N/A	47%	29%
Alcohol-related cirrhosis without HCC (CIRR)						
Total	1,653	55 [10][†]	72[†]	25.9 [4.9][†]	16%[†]	61%[†]
Germany	1,050	56 [10]	72	26.2 [5.2]	22%	61%
Switzerland	192	56 [10]	73	26.2 [5.8]	29%	48%
United Kingdom*	376	53 [11]	68	24.6 [2.6]	N/A	N/A
Italy	35	54 [9]	86	N/A	24%	66%
Alcohol misusers (ALC)						
Total	2,588	48 [10]	84	24.7 [4.1]	4%	79%
Germany	1,827	48 [9]	88	24.8 [4.3]	4%	81%
Switzerland	417	45 [12]	74	24.4 [4.0]	14%	59%
United Kingdom	344	49 [10]	76	24.7 [2.3]	2%	N/A
Italy	N/A	N/A	N/A	N/A	N/A	N/A

Bold numbers (n) indicate total n for each cohort (HCC, CIRR, ALC).

*Significance of the difference between patients with alcohol-related cirrhosis with and without HCC: $P < 0.001$.

[†]Significance of the difference between patients with alcohol-related cirrhosis and alcohol misusers: $P < 0.001$.

[‡]People with a BMI > 30 and/or with type 2 diabetes were excluded *a priori*.

Abbreviations: BMI, body mass index; T2DM, type 2 diabetes; N/A, not available.

TABLE 2. Univariate and Multivariate Analyses for Association of *HSD17B13* rs72613567 with Alcohol-related Cirrhosis and HCC in the Total European Cohort

Cohorts	HSD17B13 (rs72613567)		Comparative Groups		Genotypic OR (95% CI) Regression Model: A	Genotypic OR (95% CI) Regression Model: B	Regression Model	Allelic OR (95% CI)	Significance (P)	Cases/Controls (n)
	T T	T TA	HCC	CIRR						
Alcoholic-related cirrhosis and HCC (HCC)	T T		705	1,027			A	0.77 (0.68-0.89)	2.27 × 10 ⁻⁴	1,031/1,653
	T TA		287	524	0.80 (0.67-0.95)	0.92 (0.69-1.22)	B	0.79 (0.63-0.99)	0.037	604/679
	TA TA		39	102	0.56 (0.38-0.82)	0.43 (0.23-0.80)				
	MAF		0.177	0.220	P = 1.05 × 10 ⁻³	P = 0.028				
Alcohol-related cirrhosis without HCC (CIRR)	T T		CIRR	ALC			A	0.79 (0.63-0.98)	8.13 × 10 ⁻⁶	1,653/2,588
	T TA		1,027	1,401	0.71 (0.62-0.81)	0.79 (0.63-0.98)	B	0.85 (0.72-0.99)	0.048	679/1,483
	TA TA		524	1,009	0.78 (0.61-1.01)	0.82 (0.54-1.23)				
	MAF		0.220	0.264	P = 1.54 × 10 ⁻⁶	P = 0.085				
Alcohol misusers (ALC)	T T		HCC	ALC			A	0.60 (0.53-0.69)	1.69 × 10 ⁻¹⁴	10,31/2,588
	T TA		705	1,401	0.57 (0.48-0.66)	0.68 (0.52-0.89)	B	0.66 (0.53-0.81)	1.14 × 10 ⁻⁴	604/1,483
	TA TA		287	1,009	0.44 (0.30-0.62)	0.39 (0.21-0.72)				
	MAF		0.177	0.264	P = 3.32 × 10 ⁻¹⁴	P = 6.00 × 10 ⁻⁴				

Abbreviations: CI, confidence intervals; MAF, minor allele frequency; OR, odds ratio. Genotypic and allelic ORs were assessed by logistic regression models, model A: univariate logistic regression and model B: multivariate logistic regression, adjusted for age, sex, BMI, type 2 diabetes, and country.

viz., ALC (23.9%), CIRR (35.4%), and HCC (49.8%; Supporting Table S2).

In the univariate model, allelic and genotypic associations for *HSD17B13* rs72613567:TA were highly significant for the comparisons HCC versus CIRR ($P_{\text{allelic}} = 2.27 \times 10^{-4}$, $P_{\text{genotypic}} = 1.05 \times 10^{-3}$), CIRR versus ALC ($P_{\text{allelic}} = 8.13 \times 10^{-6}$, $P_{\text{genotypic}} = 1.54 \times 10^{-6}$), and HCC versus ALC ($P_{\text{allelic}} = 1.69 \times 10^{-14}$, $P_{\text{genotypic}} = 3.22 \times 10^{-14}$; Table 2). The protective effect associated with carriage of *HSD17B13* rs72613567:TA remained significant in the CIRR and HCC cohorts after correction for sex, age, BMI, type 2 diabetes, and country (Table 2). The protective effect for HCC was greater for homozygous than heterozygous carriage of the *HSD17B13* rs72613567:TA allele (Fig. 1A,B).

Allelic and genotypic associations for *PNPLA3* rs738409:G were also significantly associated in the CIRR ($OR_{\text{allelic}} 1.70 [1.54-1.88]$, $P_{\text{allelic}} = 1.52 \times 10^{-26}$) and HCC ($OR_{\text{allelic}} 1.77 [1.58-1.98]$, $P_{\text{allelic}} = 2.31 \times 10^{-23}$) cohorts (Table 3). These associations were robust to corrections for sex, age, BMI, type 2 diabetes, and country.

Because patients were recruited from across Europe, a meta-analysis of the *HSD17B13* and *PNPLA3* loci for association with cirrhosis and HCC was performed by country (Supporting Tables S3 and S4). The associations were not necessarily significant in every European cohort but fixed-effect meta-analyses confirmed both the protective and risk-enhancing effects of the *HSD17B13* and *PNPLA3* loci, respectively, in the total cohort. There was no evidence of heterogeneity (Supporting Figs. S1A-C and S2A-C).

The PAF for *PNPLA3* rs738409 for CIRR was 27.2% (20.7%-33.6%) and for *HSD17B13* rs72613567 -14.7% (-21.2% to -7.9%). The combined PAF for CIRR was 16.5%. The PAF% for *PNPLA3* rs738409 in HCC was 38.7% (29.3%-47.2%) and for *HSD17B13* rs72613567 -10.0% (-16.7% to -2.7%). The combined PAF for HCC was 32.6%.

The potential modifying effect of *HSD17B13* rs72613567 on *PNPLA3* rs738409 was explored by calculating the proportion of the HCC, CIRR, and ALC cohorts with different "gene signatures," combining either *PNPLA3* rs739409:G (high risk variant) with wild-type *HSD17B13* rs72613567:T (lack of protection) or the *PNPLA3* rs739409 C (low-risk variant) with *HSD17B13* rs72613567:TA (protection). A preponderance of patients in the CIRR and HCC cohorts

were homozygous for *PNPLA3* rs739409:G and for *HSD17B13* rs72613567:T, which is in line with the estimated risk contributions of each variant (Fig. 2A,B).

Possible SNP*SNP interactions between *HSD17B13* rs72613567 and *PNPLA3* rs739409 were explored by testing interaction terms in the logistic regression models for the risk for developing CIRR and the further risk for developing HCC. The genotypic interactions for the risk association with CIRR was not significant in the total study population ($P = 0.598$; Supporting Table S5) or in the men-only cohort ($P = 0.689$; Supporting Table S6). None of the *HSD17B13* \times *PNPLA3* genotype interaction pair combinations was significant. The interactions in the additive CIRR and HCC risk models were also not significant. Thus, there does not appear to be an *HSD17B13* rs72613567:TA \times *PNPLA3* rs739409:G interaction.

In a sex-specific *post hoc* analysis of the *HSD17B13* locus, an association with HCC disease risk was observed in men ($OR_{\text{allelic}} 0.75$; 95% CI, 0.64-0.87; $P = 1.72 \times 10^{-4}$) but not in women ($OR_{\text{allelic}} 1.07$; 95% CI, 0.73-1.58; $P = 0.772$), whereas the protective effect for cirrhosis risk associated with carriage of *HSD17B13* rs72613567:TA was observed in both sexes (Supporting Table S7; Fig. 3A-C). The association between carriage of *PNPLA3* rs739409:G and cirrhosis and HCC disease risk showed no sex differential (Supporting Table S8).

Discussion

The genetic contribution to the development of significant alcohol-related liver injury is gradually being unraveled. Three genetic loci have been identified that are associated with an increased risk for developing cirrhosis at genome-wide significance, namely *PNPLA3*, *TM6SF2*, *MBOAT7*,⁽¹⁹⁾ and at least two variants, *PNPLA3* rs738409 and *TM6SF2* rs58542926, are associated with an increased risk for developing HCC on this background.^(15,16,18)

There is little published evidence for genetic variants conferring protection against the development of significant alcohol-related liver injury. Indirect protection is afforded in individuals, primarily those of East Asian ancestry, who carry rs1229984 in alcohol dehydrogenase 1B⁽²⁸⁾ and/or rs671 in aldehyde dehydrogenase 2⁽²⁹⁾ and, as a consequence, develop a reaction to alcohol and so tend to avoid it.

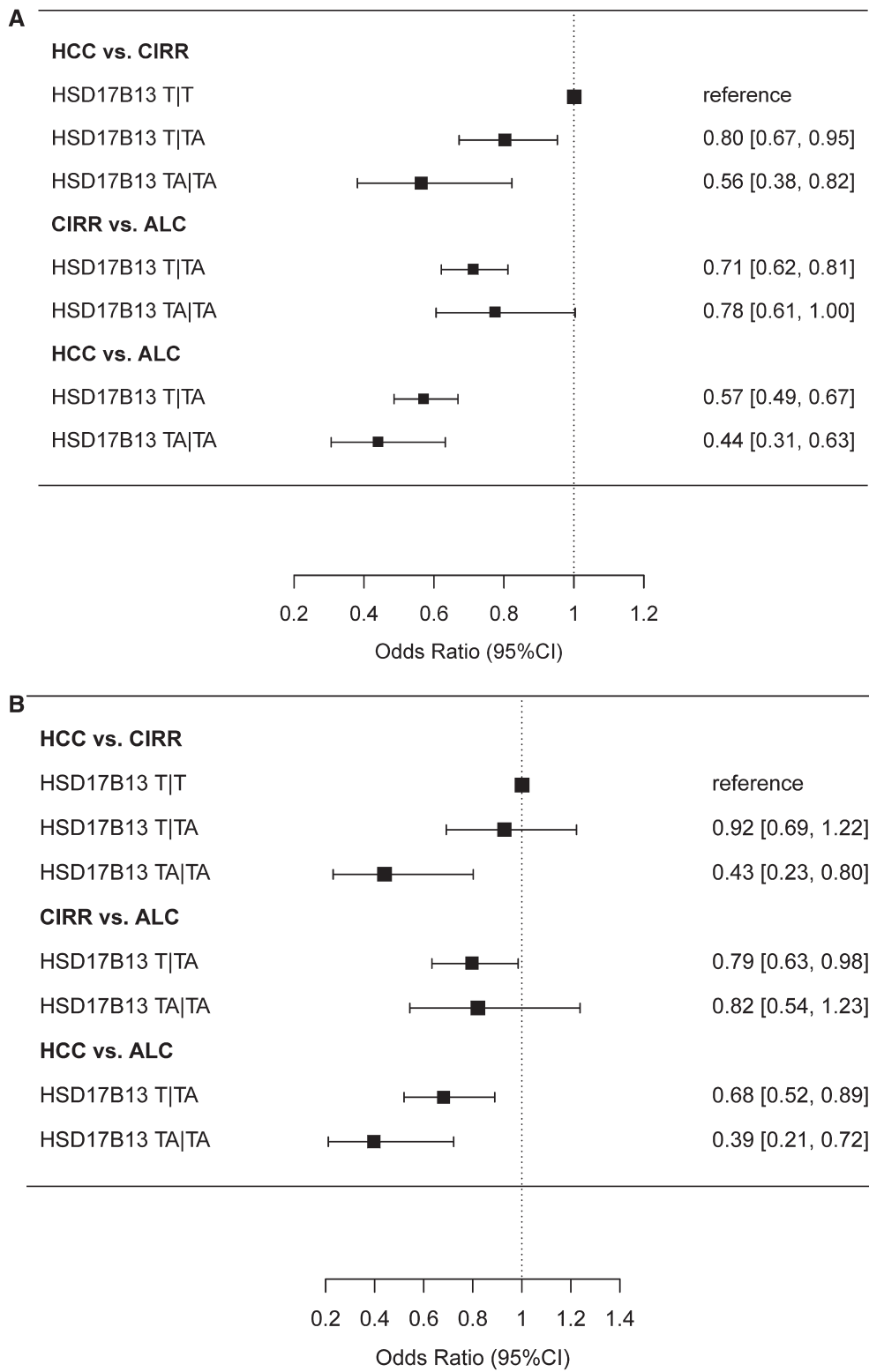


FIG. 1. Forrest plot showing genotypic ORs for heterozygous and homozygous carriage of the *HSD17B13* rs72613567:TA allele. (A) The unadjusted analysis; (B) analysis adjusted for age, sex, BMI, type 2 diabetes, and country. Homozygous carriage of *HSD17B13* rs72613567:T wild type is set as a reference.

TABLE 3. Univariate and Multivariate Analyses for Association of *PNPLA3* rs738409 with Alcohol-related Cirrhosis and HCC in the Total European Cohort

Cohorts	<i>PNPLA3</i> (rs738409)	Comparative Groups		Genotypic OR (95% CI) Regression Model: A		Genotypic OR (95% CI) Regression Model: B		Regression Model	Allelic OR (95% CI)	Significance (<i>P</i>)	Cases/Controls (<i>n</i>)
		HCC	CIRR								
Alcohol-related cirrhosis with HCC (HCC)	C C	269	697					A	1.77 (1.58-1.98)	2.31×10^{-23}	1,030/1,631
	C G	497	714	1.80 (1.50-2.16)		1.75 (1.30-2.36)		B	1.67 (1.39-2.01)	7.19×10^{-8}	603/660
	G G	264	220	3.11 (2.48-3.90)		2.75 (1.88-4.01)					
	MAF	0.498	0.354	$P = 2.98 \times 10^{-22}$		$P = 4.37 \times 10^{-7}$					
Alcohol-related cirrhosis without HCC (CIRR)	C C	697	1,364					A	1.70 (1.54-1.88)	1.52×10^{-26}	1,631/2,319
	C G	714	802	1.74 (1.52-2.00)		1.64 (1.31-2.06)		B	1.63 (1.39-1.91)	1.67×10^{-9}	660/1,469
	G G	220	153	2.81 (2.25-3.53)		2.65 (1.85-3.78)					
	MAF	0.354	0.239	$P = 1.61 \times 10^{-25}$		$P = 1.31 \times 10^{-8}$					
Alcohol misuse (ALC)	C C	269	1,364					A	3.00 (2.67-3.36)	2.17×10^{-79}	1,030/2,319
	C G	497	802	3.14 (2.65-3.73)		2.75 (2.06-3.66)		B	2.63 (2.18-3.17)	3.89×10^{-24}	603/1,469
	G G	264	153	8.75 (6.90-11.10)		6.76 (4.60-9.95)					
	MAF	0.498	0.239	$P = 4.14 \times 10^{-78}$		$P = 5.50 \times 10^{-23}$					

Abbreviations: CI, confidence intervals; MAF, minor allele frequency; OR, odds ratio Genotypic and allelic ORs were assessed by logistic regression models, model A: univariate logistic regression and model B: multivariate logistic regression, adjusted for age, sex, BMI, type 2 diabetes, and country.

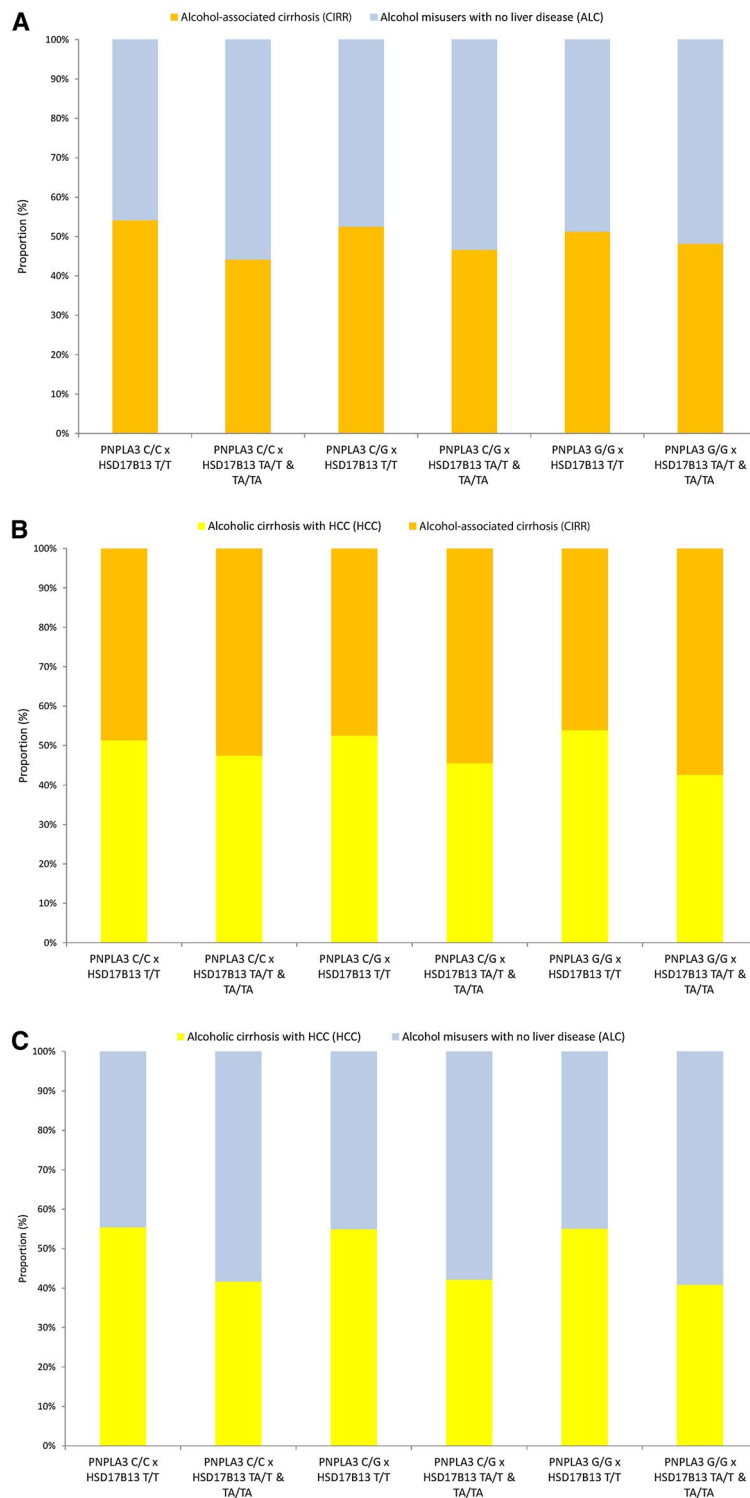


FIG. 2. (A) Relative proportions of patients with alcohol-related cirrhosis (CIRR) according to gene signature comprised of *HSD17B13* rs72613567 and *PNPLA3* rs739409 genotypes in relation to alcohol misusers without liver disease (ALC). (B) Relative proportions of patients with HCC according to gene signature comprised of *HSD17B13* rs72613567 and *PNPLA3* rs739409 genotypes in relation to patients with alcohol-related cirrhosis (CIRR). (C) Proportions of patients with HCC according to gene signature comprised of *HSD17B13* rs72613567 and *PNPLA3* rs739409 genotypes in relation to alcohol misusers with no liver disease (ALC).

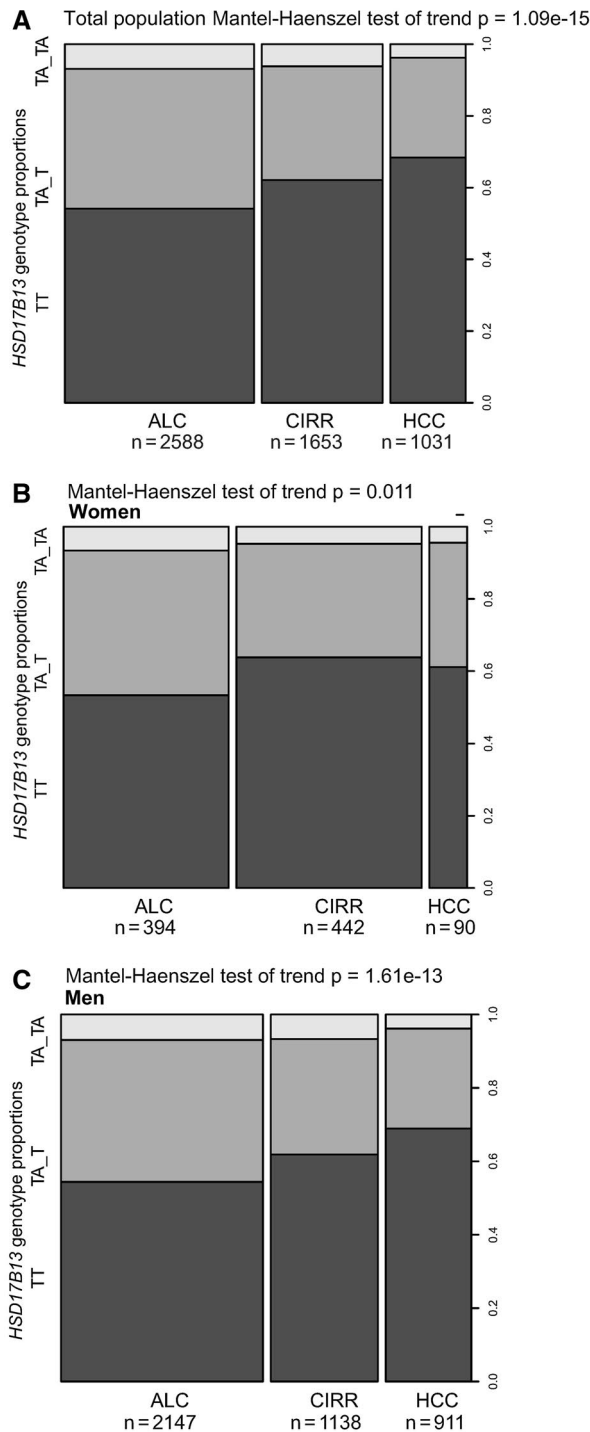


FIG. 3. (A) Test for trend on the proportion of *HSD17B13* rs72613567 genotypes in both men and women in the 3 cohorts showing a gradual decrease in TA allele carriage with escalating severity of liver disease. (B) Test for trend on the proportion of *HSD17B13* rs72613567 genotypes in women showing a decrease in TA carriage in patients with cirrhosis but not in patients with HCC. (C) Test for trend on the proportion of *HSD17B13* rs72613567 genotypes in men showing a gradual decrease of TA allele carriage with escalating severity of liver disease.

Recently, however, Abul-Husn et al.,⁽²⁰⁾ identified a splice variant, rs72613567 in *HSD17B13*, which appeared to protect against the development of chronic liver injury in people of European ancestry. Specifically, this SNP appeared to confer protection against the development of alcohol-related and NAFLD-related cirrhosis.⁽²⁰⁾ The association with alcohol-related cirrhosis was more compelling even though the total number of cases was very small, amounting to only 124 in their discovery cohort and 215 in their validation cohort.

In a later French/Belgian collaboration, Yang et al.,⁽²¹⁾ confirmed that rs72613567 in *HSD17B13* confers protection against the development of alcohol-, NAFLD-, and HCV-related liver disease. They further showed that this SNP confers protection against the development of HCC, but only when arising on a background of alcohol-related liver disease. The data were adjusted for age, sex, and the degree of hepatic fibrosis but not for other important confounders, such as BMI, type 2 diabetes, and *PNPLA3* genotype. Only half the patients with alcohol-related HCC in this study had cirrhosis. Thus, the numbers of patients with alcohol-related cirrhosis ($n = 1,243$) and alcohol-related HCC ($n = 217$) were relatively small, and those with HCC were more frequently male and older than those with cirrhosis, *per se*. Adjustments were not made for these possible confounders, leading the authors to conclude that larger cohorts were needed to validate their findings.

The cohorts included in the present study were large, consisting of 1,031 patients of European ancestry with alcohol-related cirrhosis and HCC and 1,653 with alcohol-related cirrhosis without HCC. In addition, 2,588 alcohol misusers with no evidence of liver disease were included to exclude the possibility that any observed genetic effect might relate to the susceptibility to develop problematic drinking.

The results of the present study robustly establish that carriage of *HSD17B13*:rs72613567:TA protects against the development of alcohol-related cirrhosis and its subsequent evolution to HCC. The results also reconfirm that carriage of *PNPLA3* rs738409 is associated with a significantly increased risk of developing alcohol-associated cirrhosis and HCC. However, the magnitude of these two effects is of a different order. Thus, although carriage of the *HSD17B13*:rs72613567:TA allele reduces the PAF for cirrhosis by 14.7% and for HCC by 10%, carriage of the *PNPLA3* rs738409:G

allele increases the PAFs by 27.2% and 38.7%, respectively. Consequently, the combined PAF for cirrhosis was 16.5% and for HCC 32.5%.

Thus, the risk of carriage of *PNPLA3* rs738409:G appears to be attenuated, to a degree, by co-carriage of *HSD17B13*:rs72613567:TA. Abul-Husn et al.,⁽²⁰⁾ explored the possibility of interactions between these two SNPs and observed nominally significant interactions in association analyses with serum aminotransferase activities, primarily in the obese, but there were no significant interactions in relation to chronic liver disease. They also showed, using RNA sequencing-based expression analysis, that *HSD17B13* rs72613567:TA was associated with decreased *PNPLA3* messenger RNA expression in an allele dose-dependent manner. However, exploration of SNP*SNP interaction in the present study showed no evidence of a significant interaction. Yang et al.,⁽²¹⁾ did not report data on possible SNP*SNP interaction, so no further comparisons can be made.

Male sex is a significant risk factor for the development of HCC in people with alcohol-related cirrhosis.^(12,13) The reasons for this are unknown. Men are more likely to drink at harmful levels and hence more likely to develop alcohol-related cirrhosis, but they are still proportionately overrepresented among those developing HCC. Malignant transformation is more likely to occur in individuals with alcohol-related cirrhosis who stop drinking,⁽³⁰⁾ hence the suggestion that the difference in sex-specific HCC rates may reflect lower abstinence rates among women. The possibility of sex-genetic variant interactions should also be considered. The comparative survival advantage in women with alcohol-related cirrhosis is well documented,⁽³¹⁻³³⁾ and it has recently been shown that this may relate, at least in part, to a sex-variant interaction with rs738409:G in *PNPLA3*.^(34,35)

The possibility of sex-variant interactions was also explored in the present study. The progressive increase in the risk for developing alcohol-related cirrhosis and HCC associated with carriage of *PNPLA3* rs738409:G was observed in both men and women. Likewise, the protective effect for cirrhosis risk associated with carriage of *HSD17B13* rs72613567:TA was observed in both sexes. However, the protective association of *HSD17B13* rs72613567:TA and HCC was only found in men. There was no significant sex difference in the MAFs for *HSD17B13* rs72613567:TA in the alcohol misusers (26.3% vs. 26.6%) or in the patients with alcohol-related cirrhosis (22.4% vs. 20.5%). However,

whereas the frequency was further reduced in men with HCC (17.8%), the trend partially reversed in women (21.7%). These findings are counterintuitive and are without explanation. The number of women with HCC was relatively small, so this finding needs further exploration in a larger cohort.

Neither of the previous studies explored sex differences in the effects of *HSD17B13* rs72613567:TA.^(20,21) However, Ferenci et al.,⁽³⁶⁾ found sex-related phenotypic variation associated with carriage of *HSD17B13* rs72613567:TA in patients with Wilson's disease. Thus, they showed that none of the men who developed fulminant Wilson's disease carried the protective *HSD17B13* variant in contrast to 13.6% of their female counterparts. Clearly, sex-variant interactions should be considered in any further genetic studies in the field of liver disease.

HSD17B13 belongs to the family of pluripotent 17-hydroxysteroid dehydrogenase enzymes whose members convert 17-keto- and 17-hydroxysteroids; regulate the biological activity of sex hormones; participate in fatty acid and cholesterol metabolism; and contribute to bile acid synthesis.⁽³⁷⁾ The function of *HSD17B13* is incompletely understood, but it is located on the surface of lipid droplets and is mainly expressed in the liver. Su et al.,⁽³⁸⁾ have demonstrated that transcriptional regulation of *HSD17B13* expression is likely to be liver X receptor mediated through a sterol regulatory element binding protein 1c (SREBP-1c)-dependent mechanism; they also found evidence for a SREBP-1c response element in the promoter region of the *HSD17B13* gene located on chromosome 4q22.1. *HSD17B13* expression is increased in patients with NAFLD^(39,40) and in murine models of fatty liver disease,⁽⁴¹⁾ and its overexpression in Huh7 and HepG2 hepatoma cell lines results in accumulation of *HSD17B13* on the surface of lipid droplets.^(40,42)

Ma et al.,⁽⁴³⁾ have recently shown that *HSD17B13* is an hepatic retinol dehydrogenase. Retinol, retinoic acid, and retinol-binding protein have been implicated in the pathogenesis of steatosis, fibrosis, adipogenesis, and insulin resistance.⁽⁴⁴⁾ Thus, *HSD17B13* may be involved in the complex nuclear receptor interaction in NAFLD through activation of the retinoic acid receptor.⁽⁴⁵⁾ Of greater importance, in the context of the present study, is the possibility that *HSD17B13*, functioning as a retinol dehydrogenase, may contribute to the depletion of hepatic retinoic acid observed in individuals chronically misusing

alcohol. Retinoic acid depletion results in a functional down-regulation of liver retinoic acid receptors and a marked increase in the expression of the activator protein-1 (c-Jun and c-Fos) transcriptional complex, which is associated with hepatic cell hyperproliferation, a decrease in apoptosis, and stimulated hepatic carcinogenesis.^(46,47) The retinol dehydrogenase activity of the rs72613567:TA variant is likely reduced or absent, and this may explain, at least in part, its protective effect against HCC in the context of alcohol-related cirrhosis. Accordingly, repletion of hepatic retinoic acid concentrations experimentally or in patients with HCC may have a therapeutic role.⁽⁴⁸⁾

There are some contrary findings in relation to the protective effect of a reduction in HSD17B13 activity. Chen et al.,⁽⁴⁹⁾ for example, reported that mRNA expression of HSD17B13 was down-regulated in Asians with hepatitis B virus (HBV)-related HCC, although not in HBV-related cirrhosis, and showed that low HSD17B13 expression in peritumor tissue was independently associated with a reduction in recurrence-free survival. They also showed that overexpression of HSD17B13 in Huh7 cell and SK-HEP-1 cell lines results in delays in cell cycle progression. Their overall conclusion, based on these findings, was that increased HSD17B13 expression might inhibit the development and progression of HBV-related HCC. This contrasts with the findings in the present study and those of others^(20,21) that in Europeans, decreased expression of HSD17B13 protects against the development of both alcohol-related cirrhosis and its evolution to HCC. This apparent contradiction may attest to differences in the mechanisms of HCC development in viral- and alcohol-related liver disease. Clearly, further research on the functional role of HSD17B13 in HCC development and progression is needed.

There is considerable interest in the possibility of pharmaceutical inhibition of HSD17B13 to counteract the steatogenic effect of overexpressed HSD17B13 and mimic the loss-of-function derived from the *HSD17B13* rs72613567:TA variant. For example, fenofibrate, which is an agonist of the peroxisome proliferator-activated receptor- α (PPAR α) and approved for use in humans, suppresses hepatic HSD17B13 expression in mouse liver,⁽⁵⁰⁾ a finding supported by the high expression of HSD17B13 observed in *ppara* knock-out mice.⁽⁴²⁾

This study has a number of strengths. The study cohorts were large and well-characterized and specifically selected to explore the genetic risks associated

with the development of alcohol-related cirrhosis and HCC in Caucasians of European origin. Controls were exercised for a number of known risk factors, such as age, sex, BMI, and type 2 diabetes. Similar effect sizes were observed across all individual geographical cohorts, excluding the possibility of heterogeneity and thus broadening the applicability of the findings. The specificity of the genetic associations was explored by including a large control cohort of alcohol misusers who had no evidence of liver disease.

This study also has its limitations; first, it was cross sectional and undertaken retrospectively, meaning that phenotypic data sets were not always complete. Second, it is likely that some of the patients with alcohol-related cirrhosis will develop HCC over time and equally likely that some of the individuals misusing alcohol will develop cirrhosis in the future, particularly if they continue to drink. However, the group differences were significant and remained robust to adjustment for confounders, including age. Finally, the findings cannot be used to define the genetic risks associated with the development of cirrhosis and HCC in people with liver disease of different etiologies or from outside of Europe, as evidenced by the findings of Chen and colleagues.⁽⁴⁹⁾

In conclusion: *HSD17B13* rs72613567:TA protects against the development of alcohol-related cirrhosis and, at least in men, the subsequent development of HCC. Its carriage attenuates the increased risk associated with carriage of *PNPLA3* rs738409:G. Combining phenotypic and genetic signatures to score risk could facilitate management of patients with alcohol-related liver disease. Further evaluation of the function of HSD17B13 and the rs72613567 variant may identify suitable drug targets.

Acknowledgment: The authors thank the Clinical Research Support Service of the CHUV-UNIL, Lausanne, Switzerland, for providing the infrastructure for patient recruitment and collecting phenotypic data.

Author Contributions: F.S., J.H., and M.Y.M. conceived the study; F.S. raised the pivotal funding and wrote the manuscript; M.Y.M. cowrote the manuscript and reanalyzed the data; A.M. contributed biological material from healthy controls and reviewed the biostatistics analysis; F.S., P.L., and H.D.N. contributed biological material from cases and controls; I.S., V.R., J.F., K.H.W., D.G., J.R., A.M., M.E., M.K., M.C., F.L., U.S., F.E., A.V., S.M., J.v.F., H.W., R.S., S.A., A.F., S.N., V.M., C.S., L.S., C.L., R.E.S., A.C., A.L., L.V.,

J.I.G., G.P.A., J.U.M., W.F., S.Z., J.-F.D., J.T., C.D., P.D., S.M., T.B., and M.Y.M. contributed biological material from phenotyped cases and controls; J.H. provided the genotyping infrastructure; T.B. genotyped the healthy control samples collected in the United Kingdom; S.B. performed the biostatistics and analyzed the primary data; F.S., P.L., and H.D.N. analyzed the data; P.L., S.B., H.D.N., I.S., V.R., J.F., K.H.W., D.G., J.R., A.M., M.E., M.K., M.C., F.L., T.B., A.M., U.S., F.E., A.V., S.M., J.v.F., H.W., R.S., S.A., A.F., S.N., V.M., C.S., L.S., C.L., R.E.S., A.C., A.L., L.V., J.I.G., G.P.A., J.U.M., W.F., S.Z., J.-F.D., J.T., C.D., P.D., S.M., T.B., J.H., and M.Y.M. critically revised the manuscript; F.S., P.L., S.B., H.D.N., I.S., V.R., J.F., K.H.W., D.G., J.R., A.M., M.E., M.K., M.C., F.L., T.B., A.M., U.S., F.E., A.V., S.M., J.v.F., H.W., R.S., S.A., A.F., S.N., V.M., C.S., L.S., C.L., R.E.S., A.C., A.L., L.V., J.I.G., G.P.A., J.U.M., W.F., S.Z., J.-F.D., J.T., C.D., P.D., S.M., T.B., J.H., and M.Y.M. approved the final draft submitted.

REFERENCES

- Singal AK, Bataller R, Ahn J, Ahn J, Kamath PS, Shah VH. ACG clinical guideline: alcoholic liver disease. *Am J Gastroenterology* 2018;113:175-194.
- Asrani SK, Devarbhavi H, Eaton J, Kamath PS. Burden of liver diseases in the world. *J Hepatol* 2019;70:151-171.
- Rehm J, Samokhvalov AV, Shield KD. Global burden of alcoholic liver diseases. *J Hepatol* 2013;59:160-168.
- Burra P, Senzolo M, Adam R, Delvart V, Karam V, Germani G, et al.; for ELITA and on behalf of ELTR Liver Transplant Centers. Liver transplantation for alcoholic liver disease in Europe: a study from the ELTR (European Liver Transplant Registry). *Am J Transplant* 2010;10:138-148.
- Parker R, Aithal GP, Becker U, Gleeson D, Masson S, Wyatt JI, Rowe IA; WALDO study group. Natural history of histologically proven alcohol-related liver disease: a systematic review. *J Hepatol* 2019;71:586-593.
- Leevy CM. Cirrhosis in alcoholics. *Med Clin North Am* 1968;52:1445-1455.
- Teli MR, Day CP, Burt AD, Bennett MK, James OF. Determinants of progression to cirrhosis or fibrosis in pure alcoholic fatty liver. *Lancet* 1995;346:987-990.
- Kodama K, Tokushige K, Hashimoto E, Tani M, Shiratori K. Hepatic and extrahepatic malignancies in cirrhosis caused by non-alcoholic steatohepatitis and alcoholic liver disease. *Alcohol Clin Exp Res* 2013;37:E247-E252.
- Mancebo A, González-Diéguez ML, Cadahía V, Varela M, Pérez R, Navascués CA, et al. Annual incidence of hepatocellular carcinoma among patients with alcoholic cirrhosis and identification of risk groups. *Clin Gastroenterol Hepatol* 2013;11:95-101.
- Ganne-Carrié N, Chaffaut C, Bourcier V, Archambeaud I, Perarnau JM, Oberti F, et al. Estimate of hepatocellular carcinoma incidence in patients with alcoholic cirrhosis. *J Hepatol* 2018;69:1274-1283.
- Ganne-Carrié N, Nahon P. Hepatocellular carcinoma in the setting of alcohol-related liver disease. *J Hepatol* 2019;70:284-293.
- Kim D, Li AA, Perumpail BJ, Gadiparthi C, Kim W, Cholanteril G, et al. Changing trends in etiology- and ethnicity-based annual mortality rates of cirrhosis and hepatocellular carcinoma in the United States. *HEPATOLOGY* 2019;69:1064-1074.
- Stickel F, Moreno C, Hampe J, Morgan MY. Genetics of alcohol dependence and alcohol-related liver disease. *J Hepatol* 2017;66:195-211.
- Stickel F, Buch S, Lau K, Meyer zu Schwabedissen H, Berg T, Ridinger M, et al. Genetic variation in the *PNPLA3* gene is associated with alcoholic liver injury in Caucasians. *HEPATOLOGY* 2011;53:86-95.
- Chamorro AJ, Torres JL, Mirón-Canelo JA, González-Sarmiento R, Laso FJ, Marcos M. Systematic review with meta-analysis: the I148M variant of patatin-like phospholipase domain-containing 3 gene (*PNPLA3*) is significantly associated with alcoholic liver cirrhosis. *Aliment Pharmacol Ther* 2014;40:571-581.
- Singal AG, Manjunath H, Yopp AC, Beg MS, Marrero JA, Gopal P, et al. The effect of *PNPLA3* on fibrosis progression and development of hepatocellular carcinoma: a meta-analysis. *Am J Gastroenterol* 2014;109:325-334.
- Trépo E, Nahon P, Bontempi G, Valenti L, Falletti E, Nischalke HD, et al. Association between the *PNPLA3* (rs738409 C>G) variant and hepatocellular carcinoma: evidence from a meta-analysis of individual participant data. *HEPATOLOGY* 2014;59:2170-2177.
- Stickel F, Buch S, Nischalke HD, Weiss KH, Gotthardt D, Fischer J, et al. Genetic variants in *PNPLA3* and *TM6SF2* predispose to the development of hepatocellular carcinoma in individuals with alcohol-related cirrhosis. *Am J Gastroenterol* 2018;113:1475-1483.
- Buch S, Stickel F, Trépo E, Way M, Herrmann A, Nischalke HD, et al. A two-stage genome-wide association study confirms *PNPLA3* and identifies *TM6SF2* and *MBOAT7* as novel risk loci for alcohol-related cirrhosis. *Nat Genet* 2015;47:1443-1448.
- Abul-Husn NS, Cheng X, Li AH, Xin Y, Schurmann C, Stevis P, et al. A protein-truncating *HSD17B13* variant and protection from chronic liver disease. *N Engl J Med* 2018;378:1096-1106.
- Yang J, Trépo E, Nahon P, Cao Q, Moreno C, Letouze E, et al. A 17-beta-hydroxysteroid dehydrogenase 13 variant protects from hepatocellular carcinoma development in alcoholic liver disease. *HEPATOLOGY* 2019;70:231-240.
- European Association for the Study of the Liver. *EASL Clinical Practice Guidelines: management of hepatocellular carcinoma*. *J Hepatol* 2018;69:182-236.
- Pugh RN, Murray-Lyon IM, Dawson JL, Pietroni MC, Williams R. Transection of the oesophagus for bleeding oesophageal varices. *Br J Surg* 1973;60:646-649.
- American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*. 4th ed. Washington, DC: American Psychiatric Press; 1994.
- Witte JS, Visscher PM, Wray NR. The contribution of genetic variants to disease depends on the ruler. *Nat Rev Genet* 2014;15:765-776.
- Anderson CA, Pettersson FH, Clarke GM, Cardon LR, Morris AP, Zondervan KT. Data quality control in genetic case-control association studies. *Nat Protoc* 2010;5:1564-1573.
- 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, et al. A global reference for human genetic variation. *Nature* 2015;526:68-74.
- Li D, Zhao H, Gelernter J. Strong association of the alcohol dehydrogenase 1B gene (*ADH1B*) with alcohol dependence and alcohol-induced medical diseases. *Biol Psychiatry* 2011;70:504-512.
- Li D, Zhao H, Gelernter J. Strong protective effect of the aldehyde dehydrogenase gene (*ALDH2*) 504lys (*2) allele against alcoholism and alcohol-induced medical diseases in Asians. *Hum Genet* 2012;131:725-737.
- Lee FI. Cirrhosis and hepatoma in alcoholics. *Gut* 1966;7:77-85.

- 31) Schlichting P, Christensen E, Andersen PK, Fauerholdt L, Juhl E, Poulsen H, et al. Prognostic factors in cirrhosis identified by Cox's regression model. *HEPATOLOGY* 1983;3:889-995.
- 32) D'Amico G, Morabito A, Pagliaro L, Marubini E. Survival and prognostic indicators in compensated and decompensated cirrhosis. *Dig Dis Sci* 1986;31:468-475.
- 33) Ginés P, Quintero E, Arroyo V, Terés J, Bruguera M, Rimola A, et al. Compensated cirrhosis: natural history and prognostic factors. *HEPATOLOGY* 1987;7:122-128.
- 34) Meffert PJ, Repp KD, Völzke H, Weiss FU, Homuth G, Kühn JP, et al. The *PNPLA3* SNP rs738409: G allele is associated with increased liver disease-associated mortality but reduced overall mortality in a population-based cohort. *J Hepatol* 2018;68:858-860.
- 35) Atkinson SR, Way MJ, McQuillin A, Morgan MY, Thursz MR. Reply to: "The *PNPLA3* SNP rs738409: G allele is associated with increased liver disease-associated mortality but reduced overall mortality in a population-based cohort". *J Hepatol* 2018;68:860-862.
- 36) Ferenci P, Pfeifferberger J, Stättermayer AF, Stauber RE, Willheim C, Weiss KH, et al. *HSD17B13* truncated variant is associated with a mild hepatic phenotype in Wilson's Disease. *JHEP Rep* 2019;1:2-8.
- 37) Saloniemi T, Jokela H, Strauss L, Pakarinen P, Poutanen M. The diversity of sex steroid action: novel functions of hydroxysteroid (17 β) dehydrogenases as revealed by genetically modified mouse models. *J Endocrinol* 2012;212:27-40.
- 38) Su W, Peng J, Li S, Dai YB, Wang CJ, Xu H, et al. Liver X receptor α induces 17 β -hydroxysteroid dehydrogenase-13 expression through SREBP-1c. *Am J Physiol Endocrinol Metab* 2017;312:E357-E367.
- 39) Su W, Wang Y, Jia X, Wu W, Li L, Tian X, et al. Comparative proteomic study reveals 17 β -HSD13 as a pathogenic protein in nonalcoholic fatty liver disease. *Proc Natl Acad Sci USA* 2014;111:11437-11442.
- 40) Kampf C, Mardinoglu A, Fagerberg L, Hallström BM, Edlund K, Lundberg E, et al. The human liver-specific proteome defined by transcriptomics and antibody-based profiling. *FASEB J* 2014;28:2901-2914.
- 41) Adam M, Heikelä H, Sobolewski C, Portius D, Mäki-Jouppila J, Mehmood A, et al. Hydroxysteroid (17 β) dehydrogenase 13 deficiency triggers hepatic steatosis and inflammation in mice. *FASEB J* 2018;32:3434-3447.
- 42) Horiguchi Y, Araki M, Motojima K. 17 β -Hydroxysteroid dehydrogenase type 13 is a liver-specific lipid droplet-associated protein. *Biochem Biophys Res Commun* 2008;370:235-238.
- 43) Ma Y, Belyaeva OV, Brown PM, Fujita K, Valles K, Karki S, et al. 17 β -hydroxysteroid dehydrogenase 13 is a hepatic retinol dehydrogenase associated with histological features of nonalcoholic fatty liver disease. *HEPATOLOGY* 2019;69:1504-1519.
- 44) Saeed A, Dullaart RPF, Schreuder TCMA, Blokzijl H, Faber KN. Disturbed vitamin A metabolism in non-alcoholic fatty liver disease (NAFLD). *Nutrients* 2017;10:29.
- 45) Kim SC, Kim CK, Axe D, Cook A, Lee M, Li T, et al. All-trans-retinoic acid ameliorates hepatic steatosis in mice by a novel transcriptional cascade. *HEPATOLOGY* 2014;59:1750-1760.
- 46) Wang XD, Liu C, Chung J, Stickel F, Seitz HK, Russell RM. Chronic alcohol intake reduces retinoic acid concentration and enhances AP-1 (c-Jun and c-Fos) expression in rat liver. *HEPATOLOGY* 1998;28:744-750.
- 47) Cortes E, Lachowski D, Rice A, Chronopoulos A, Robinson B, Thorpe S, et al. Retinoic acid receptor- β is downregulated in hepatocellular carcinoma and cirrhosis and its expression inhibits myosin-driven activation and durotaxis in hepatic stellate cells. *HEPATOLOGY* 2019;69:785-802.
- 48) Muto Y, Moriwaki H, Saito A. Prevention of second primary tumors by an acyclic retinoid in patients with hepatocellular carcinoma. *N Engl J Med* 1999;340:1046-1047.
- 49) Chen J, Zhuo JY, Yang F, Liu ZK, Zhou L, Xie HY, et al. 17 β -hydroxysteroid dehydrogenase 13 inhibits the progression and recurrence of hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int* 2018;17:220-226.
- 50) Rotroff DM, Pijut SS, Marvel SW, Jack JR, Havener TM, Pujol A, et al. ACCORD/ACCORDion Investigators. Genetic variants in *HSD17B3*, *SMAD3*, and *IPO11* impact circulating lipids in response to fenofibrate in individuals with type 2 diabetes. *Clin Pharmacol Ther* 2018;103:712-721.

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