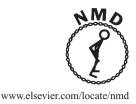




Available online at www.sciencedirect.com





Neuromuscular Disorders 27 (2017) 1043-1046

Case report

A homozygous *DPM3* mutation in a patient with alpha-dystroglycan-related limb girdle muscular dystrophy

P.Y.K. Van den Bergh ^{a,*}, Y. Sznajer ^{a,b}, V. Van Parys ^a, W. van Tol ^{c,d}, R.A. Wevers ^d, D.J. Lefeber ^{c,d}, L. Xu ^{e,f}, M. Lek ^{e,f}, D.G. MacArthur ^{e,f}, K. Johnson ^g, L. Phillips ^g, A. Töpf ^g,

V. Straub^g

^a Neuromuscular Reference Centre, University Hospital St-Luc, University of Louvain, Brussels, Belgium

^b Centre for Human Genetics, University Hospital St-Luc, University of Louvain, Brussels, Belgium

^c Department of Neurology, Radboud University Medical Centre, Nijmegen, The Netherlands

^d Translational Metabolic Laboratory, Radboud University Medical Centre, Nijmegen, The Netherlands

° Analytic and Translational Genetics Unit, Massachusetts General Hospital, Boston, MA 02114, USA

^f Program in Medical and Population Genetics, Broad Institute of Harvard and MIT, Cambridge, MA 02142, USA

^g The John Walton Muscular Dystrophy Research Centre, Institute of Genetic Medicine, Newcastle University, Newcastle upon-Tyne, United Kingdom

Received 19 April 2017; received in revised form 27 June 2017; accepted 12 July 2017

Abstract

Defects of O-linked glycosylation of alpha-dystroglycan cause a wide spectrum of muscular dystrophies ranging from severe congenital muscular dystrophy associated with abnormal brain and eye development to mild limb girdle muscular dystrophy. We report a female patient who developed isolated pelvic girdle muscle weakness and wasting, which became symptomatic at age 42. Exome sequencing uncovered a homozygous c.131T > G (p.Leu44Pro) substitution in *DPM3*, encoding dolichol-P-mannose (DPM) synthase subunit 3, leading to a 50% reduction of enzymatic activity. Decreased availability of DPM as an essential donor substrate for protein O-mannosyltransferase (POMT) 1 and 2 explains defective skeletal muscle alpha-dystroglycan O-glycosylation. Our findings show that *DPM3* mutations may lead to an isolated and mild limb girdle muscular dystrophy phenotype without cardiomyopathy.

 $\ensuremath{\mathbb{C}}$ 2017 Elsevier B.V. All rights reserved.

Keywords: Limb girdle muscular dystrophy; Alpha-dystroglycan; Dolichol-P-mannose synthase; DPM3

1. Introduction

Limb girdle muscular dystrophies (LGMD) represent an increasingly large and heterogeneous group of autosomal dominant and recessive disorders. In many patients, the molecular origin remains unknown and next generation sequencing has become a very important tool to hasten the genetic diagnosis and to identify variants and mutations in genes not previously associated with LGMD. Here, we report an adult female patient with autosomal recessive LGMD (LGMD2), in whom exome sequencing by inclusion in the MYO-SEQ project (Newcastle upon-Tyne, UK) revealed a homozygous substitution in *DPM3*, encoding dolichol-P-mannose (DPM) synthase subunit 3.

https://doi.org/10.1016/j.nmd.2017.07.006 0960-8966/© 2017 Elsevier B.V. All rights reserved.

2. Case report

The patient's medical history was uneventful. Early motor developmental milestones normally acquired and the patient started walking at the age of 1 year. At age 30 years, the patient presented with right-sided painful brachial plexopathy. Otherwise, the clinical neurological examination was normal. The neurological work-up was indicative of an inflammatory origin and the patient was diagnosed with neuralgic amyotrophy. She recovered within a few weeks. Surprisingly, serum creatine kinase (CK) activity was elevated at 4310 IU/L (N < 200). A deltoid muscle biopsy only showed mild nonspecific myopathic changes. At age 42, the patient had difficulty rising from a chair and developed an unsteady gait with tendency to fall. Because of persistingly high CK levels (2732 IU/L), a quadriceps muscle biopsy was performed, which showed a dystrophic pattern and alpha-dystroglycan (aDG) deficiency as demonstrated by immunoblotting with a IIH6C4 antibody at 1:500 dilution (Millipore SA, Overijse, Belgium)

^{*} Corresponding author. Neuromuscular Reference Centre, University Hospital St-Luc, Avenue Hippocrate 10, 1200 Brussels, Belgium.

E-mail address: peter.vandenbergh@uclouvain.be (P.Y.K. Van den Bergh).

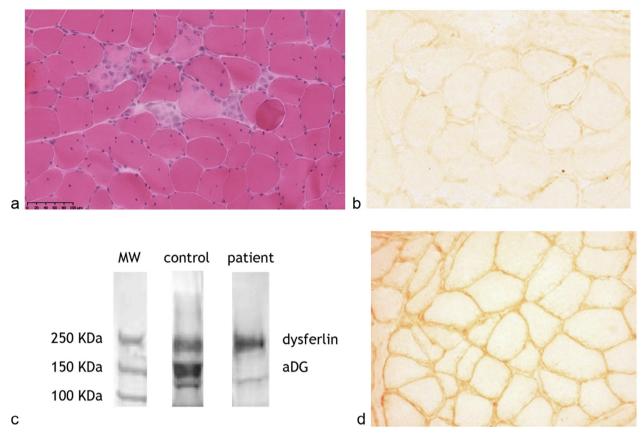


Fig. 1. Left quadriceps muscle biopsy. (A) HE showing muscle fibre necrosis, central nuclei, increased fibre size variability. aDG immunohistochemistry using IIH6C4 antibody (Millipore SA, Overijse, Belgium, DAB) showing almost complete absence of sarcolemmal staining (B) but not in a control muscle biopsy (D). (C) Western blot confirming absence of aDG (IIH6C4 antibody, 1/500 dilution).

(Fig. 1). This led to the diagnosis of aDG-related LGMD. No mutations in FKRP were found. At age 57, proximal lower limb weakness had clearly progressed. The patient could not get up from a chair without using her hands and had difficulty going upstairs. Manual muscle testing (MRC grades) showed the following abnormalities: gluteus maximus 0/5; iliopsoas and quadriceps 3/5; gluteus medius and adductors 2/5; hamstrings 4/5, tibialis anterior, tibialis posterior, peroneus longus, and triceps surae 3/5. Gowers sign was positive. Deep tendon reflexes, cranial nerves and sensation were normal. Brain MRI, respiratory, cardiac and ophthalmologic work-up were unremarkable. Nerve conduction studies were normal. EMG showed brief small amplitude polyphasic motor unit action potentials with early recruitment in the iliopsoas muscle. Muscle MRI results are shown in Fig. 2. There is no family history of neuromuscular disease. The mother died at age 94 of pancreas carcinoma. The father is healthy at age 95. One brother died at age 58 of intestinal cancer, one sister at age 39 of a brain tumour. Two other sisters and 2 sons are healthy. Serum CK levels were normal in the father and the 2 sisters.

3. Molecular analysis

Exome sequencing by inclusion into MYO-SEQ (Newcastle University, Newcastle upon-Tyne, UK) was performed at the Broad Institute's Genomics Platform, using Illumina exome

capture. A homozygous c.131T > G (p.Leu44Pro) substitution was identified in DPM3 (gene coverage 95%). DPM3 encodes DPM synthase subunit 3. This change is extremely rare in the control population (MAF = 0.00084%) and only found in the heterozygous state. Leu44 is an evolutionary highly conserved amino acid and a change to proline is predicted to be pathogenic by in silico tools (Mutation Taster, PolyPhen, UMD-Predictor and SIFT). Segregation studies identified heterozygosity in the father and in one of the two sisters. Maternal DNA was not available. Transferrin isoelectric focusing as well as mass spectroscopy of transferrin N-glycans in serum [1] were normal, indicating that N-glycosylation was well preserved in liver and serum. DPM synthase activity was analysed according to Barone et al [2] by measuring the formation of radio-active DPM in cultured fibroblasts and was reduced by 50% as compared to a healthy control.

4. Discussion

We report a patient with mild LGMD2 and without central nervous system involvement, caused by a homozygous substitution in *DPM3*. This gene encodes DPM synthase subunit 3 and we found that the enzymatic activity of DPM synthase was reduced by 50%. DMP synthase is an enzyme complex composed of 3 protein subunits, DPM1, DPM2, and DPM3. Whereas DPM2 stabilises the complex, DPM3 tethers

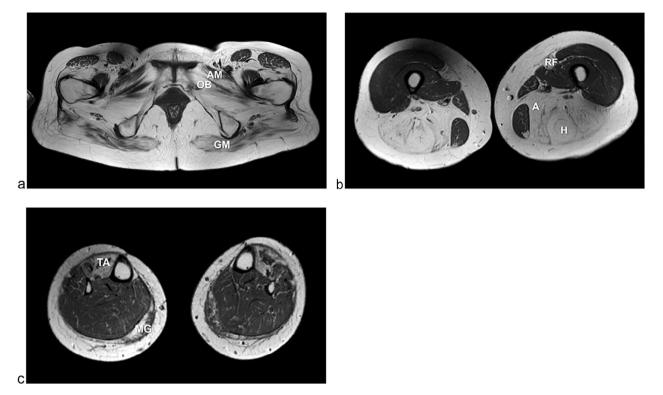


Fig. 2. T1 axial MRI sections showing increased signal intensity in (A) the gluteus maximus (GM), adductor magnus (AM) and obturatorius externus (OB) muscles, (B) in the hamstring (H), adductor (A) and rectus femoris (RF) muscles, and (C) in the tibialis anterior (TA) and medial gastrocnemius (MG) muscles.

the catalytic subunit (DPM1) to the endoplasmic reticulum membrane [3]. DPM synthase catalyses the synthesis of DPM from GDP-mannose and dolichol phosphate. As DPM is an essential donor substrate required in different glycosylation pathways (N-glycosylation, C- and O-mannosylation, GPIanchor formation) [4,5], it is not surprising that of O-mannosylation aDG. catalysed by protein O-mannosyltransferase (POMT) 1 and 2, is compromised when one of the subunit-encoding genes is mutated. This was first reported in an 11year-old female patient with mild LGMD2 and a 254T > C (p.L85S) mutation in *DPM3* [6]. At age 20, she developed dilated cardiomyopathy and at age 21, she had a stroke-like episode involving the right temporo-parietal region with normal brain MRI. In addition to abnormal N-glycosylation, deficient O-mannosylation of aDG was confirmed in a muscle biopsy and the disorder was classified as DPM3-CDG. Later, Barone et al [2] reported 3 children from 2 families with DPM2 mutations with profound developmental delay, intractable seizures, microcephaly, and early fatal outcome. The patients had aDG-deficient congenital muscular dystrophy. DPM1 mutations have been reported in 7 cases with various degrees and combinations of early onset encephalopathy, seizures, microcephaly, dysmorphic features, developmental delay, optic atrophy, and cerebellar dysfunction [7-10]. These were classified as Congenital Disorders of Glycosylation (CDG) type I (DPM1-CDG) due to abnormal N-glycosylation. In 5 of these patients, CK levels were elevated but evidence of muscular dystrophy was not reported. In 2013, Yang et al [11] reported an infant with DPM1 mutations and

showed deficient O-mannosylation of aDG as well, presenting with aDG-deficient congenital muscular dystrophy with seizures but otherwise minimal central nervous system involvement on MRI only. Our findings show that abnormal aDG O-mannosylation related to *DPM3* mutations may lead to LGMD2 phenotype without cardiomyopathy or central nervous system involvement and with presumably normal N-glycosylation.

References

- [1] van Scherpenzeel M, Steenbergen G, Morava E, Wevers RA, Lefeber DJ. High-resolution mass spectrometry glycoprofiling of intact transferrin for diagnosis and subtype identification in the congenital disorders of glycosylation. Transl Res 2015;166:639–49.
- [2] Barone R, Aiello C, Race V, Morava E; Foulquier F; Riemersma M et al. DPM2-CDG: a muscular dystrophy-dystroglycanopathy syndrome with severe epilepsy. Ann Neurol 2012;72:550–8.
- [3] Ashida H, Maeda Y, Kinoshita T. DPM1, the catalytic subunit of dolichol-phosphate mannose synthase, is tethered to and stabilized on the endoplasmic reticulum membrane by DPM3. J Biol Chem 2006;281:896–904.
- [4] Maeda Y, Tanaka S, Hino J, Kangawa K, Kinoshita T. Human dolichol-phosphate-mannose synthase consists of three subunits, DPM1, DPM2 and DPM3. EMBO J 2000;19:2475–82.
- [5] Schenk B, Fernandez F, Waechter CJ. The inside and outside of dolichyl phosphate biosynthesis and recycling in the endoplasmic reticulum. Glycobiology 2001;11:61R–70R.
- [6] Lefeber DJ, Schonberger J, Morava E, Guillard M, Huyben KM, Verrijp K et al. Deficiency of Dol-P-Man synthase subunit DPM3 bridges the congenital disorders of glycosylation with the dystroglycanopathies. Am J Hum Genet 2009;85:76–86.

- [7] Imbach T, Schenk B, Schollen E, Burdan P, Stutz A, Grunewald S et al. Deficiency of dolichol-phosphate-mannose synthase-1 causes congenital disorder of glycosylation type Ie. J Clin Invest 2000;105:233– 9.
- [8] Kim S, Westphal V, Srikrishna G, Mehta DP, Peterson S, Filiano J et al. Dolichol phosphate mannose synthase (*DPM1*) mutations define congenital disorder of glycosylation Ie (CDG-Ie). J Clin Invest 2000;105:191–8.
- [9] García-Silva MT, Matthijs G, Schollen E, Cabrera JC, Sanchez del Pozo J, Martin Herreros M et al. Congenital disorder of glycosylation

(CDG) type Ie. A new patient. J Inherit Metab Dis 2004;27:591-600.

- [10] Dancourt J, Vuillaumier-Barrot S, de Baulny HO, Sfaello I, Barnier A, Le Bizec C et al. A new intronic mutation in the *DPM1* gene is associated with a milder form of CDG Ie in two French siblings. Pediatr Res 2006;59:835–9.
- [11] Yang AC, Ng BG, Moore SA, Rush J, Waechter CJ, Raymond KM et al. Congenital disorder of glycosylation due to *DPM1* mutations presenting with dystroglycanopathy-type congenital muscular dystrophy. Mol Genet Metab 2013;110:345–51.