Modulation of seizure threshold by vagus nerve stimulation in an animal model for motor seizures

De Herdt V, De Waele J, Raedt R, Wyckhuys T, El Tahry R, Vonck K, Wadman W, Boon P. Modulation of seizure threshold by vagus nerve stimulation in an animal model for motor seizures. Acta Neurol Scand: 2010: 121: 271–276.

© 2009 The Authors Journal compilation © 2009 Blackwell Munksgaard.

Objective - The precise mechanism of action of vagus nerve stimulation (VNS) in suppressing epileptic seizures remains to be elucidated. This study investigates whether VNS modulates cortical excitability by determining the threshold for provoking focal motor seizures by cortical electrical stimulation before and after VNS. Material and *methods* – Male Wistar rats (n = 8) were implanted with a cuffelectrode around the left vagus nerve and with stimulation electrodes placed bilaterally on the rat motor cortex. Motor seizure threshold (MST) was assessed for each rat before and immediately after 1 h of VNS with standard stimulation parameters, during two to three sessions on different days. Results - An overall significant increase of the MST was observed following 1 h of VNS compared to the baseline value (1420 μ A and 1072 μ A, respectively; $\hat{P} < 0.01$). The effect was reproducible over time with an increase in MST in each experimental session. Conclusions - VNS significantly increases the MST in a cortical stimulation model for motor seizures. These data indicate that VNS is capable of modulating cortical excitability.

Introduction

Vagus nerve stimulation (VNS) is indicated in patients with refractory epilepsy who are unsuitable candidates for epilepsy surgery. By means of a helical electrode that is wound around the left vagus nerve, electrical stimuli are administered through an implantable and programmable pulse generator. The precise mechanism of action (MOA) by which VNS suppresses epileptic seizures remains to be elucidated. It has been demonstrated that VNS has both an acute effect on seizures, is able to interrupt ongoing seizure activity, as well as having a more chronic seizure preventative effect following long-term treatment (1). The acute and chronic effects are likely to be based on a distinct MOA that involves different neurochemical and neuromodulatory changes

V. De Herdt¹, J. De Waele², R. Raedt¹, T. Wyckhuys¹, R. El Tahry¹, K. Vonck¹, W. Wadman^{1,3}, P. Boon¹

¹Laboratory for Clinical and Experimental Neurophysiology, Department of Neurology, Ghent University Hospital, Ghent, Belgium; ²Intensive Care Unit, Ghent University Hospital, Ghent, Belgium; ³Swammerdam Institute of Life Sciences, Department of Neurobiology, University of Amsterdam, Amsterdam, the Netherlands

Key words: cortical excitability; cortical stimulation; seizure threshold; vagus nerve stimulation

Veerle De Herdt, Department of Neurology 1K12IA, Reference Center for Refractory Epilepsy, Ghent University Hospital, De Pintelaan 185, 9000 Ghent, Belgium

Tel.: +32 9 332 64 81 Fax: +32 9 332 45 47 e-mail: veerle.deherdt@ugent.be

Accepted for publication April 20, 2009

affecting cortical excitability. Further investigation of these distinct effects may clarify the underlying MOA of VNS, which may ultimately improve responder rates in patients, as currently, VNS treatment is unsuccessful in about one third of the treated patients (2).

In 1989, Voskuyl et al. described an animal model that allows to investigate cortical excitability (3). In this cortical stimulation model, the threshold for evoking convulsions is determined by electrical stimulation of the motor cortex in unanaesthetized rats. It is a reliable acute seizure model that allows to repeatedly test changes in seizure threshold and it has previously been used to evaluate anticonvulsant drug activity (3–7). We hypothesized that modulation of cortical excitability could be the MOA through which VNS exerts its acute anti-seizure effect.

Material and methods

Animals

Eight male Wistar rats (Harlan, the Netherlands) weighing 250–300 g, were treated according to guidelines approved by the European Ethics Committee (decree 86/609/EEC). The study protocol was approved by the Animal Experimental Ethical Committee of Ghent University Hospital (ECP 05/17). All animals were kept under environmentally controlled conditions (12 h light/dark cycles, 20–23°C and 50% relative humidity) with food and water intake *ad libitum*.

Surgical procedures

The animals were implanted with six epidural registration/stimulation electrodes and a custom made silicone spiral cuff-electrode with platinum contacts around the left vagus nerve. All rats were anaesthetized with a ketamine/xylazine (80 and 7.5 mg/kg respectively, i.p.) mixture. For implantation of the vagus nerve cuff-electrode an incision was made over the left anterior cervical region. The cuff-electrode was wound around the left vagus nerve and the ends were tunnelled under the skin over the back of the neck towards the head, where they were fixated in a head cap using acrylic cement. For stimulation of the motor cortex, two epidural stainless steel screw electrodes were positioned over the motor area of the left and right frontal cortex (3.0 mm left and right of the midline, 1.7 mm anterior to bregma). Four epidural stainless steel screw electrodes were implanted bilaterally on the parietal cortex; three of them were used for electroencephalogram (EEG) recording, the fourth was used as the reference electrode. The leads of the epidural electrodes were fixed together with the leads of the vagus nerve cuff-electrode in the head cap on the skull of the rat using acrylic cement. The animals were allowed to recover from surgery for 2 weeks before the experiments were performed.

Cortical stimulation, vagus nerve stimulation and video-EEG monitoring

The experiments were performed between 9.00 am and 01.00 pm and repeated several times for each animal on different days. In total, two to three sessions were performed for each animal, with at least 3 days in between sessions. During the experiments, the animals were connected through a multifunctional commutator: (i) to allow seizure observation and EEG recording using a digital video-EEG monitoring system, (ii) to perform cortical stimulation using a custom made external current stimulator and (iii) to perform VNS using an external current stimulator (NCP, model 100; Cyberonics Inc., Houston, TX, USA). Rats were freely moving in their cages.

At the beginning of each experiment, the impedance of the electrode-to-vagus nerve interface was measured.

One experimental session started with a baseline cortical stimulation session (baseline condition). Immediately following 1 h of VNS (0.75 mA, 30 Hz, 250 μ s, 30 s on/1.8 min off), a second cortical stimulation session was performed (VNS-condition). Stimulation of the motor cortex was performed using a ramp-shape pulse train of biphasic rectangular pulses (600 μ s, 50 Hz, 0–2000 μ A). The maximal duration of the cortical stimulation train was 15 s. The stimulation was interrupted when a focal seizure was noticed by visual inspection. During one cortical stimulation trains were given with a minimum interval of 3 min.

To determine the motor seizure threshold (MST), a *post hoc* analysis of the video images was performed by an experienced observer, without prior knowledge of the treatment (baseline or VNS condition). The MST is the current intensity corresponding to the first clinical symptoms of a focal seizure. A focal seizure is characterized by forelimb clonus, a tonic backward movement of the body or an axial myoclonic seizure. In the recorded video image a timer with a precision of 1/100 s was used to determine the exact beginning of the focal seizure. This time moment was then converted to the stimulation intensity (μA) of the ramp-shape pulse train at that exact moment. The MST during the baseline and VNS condition in each rat were calculated as the mean of at least three consecutive stimulation trains.

As an additional control to ensure that MST values did not increase over time within one experimental session, several sessions of SHAM stimulation were performed in rats, that showed the greatest increase in MST during the VNS condition. A SHAM stimulation session comprised a baseline MST determination followed by a second MST determination after 1 h without VNS.

Statistical analysis

Differences in MST were calculated parametrically using the paired Student's *t*-test (P < 0.05). Delta values were defined as the difference in MST during the baseline and VNS condition.

Results

In total, 21 experimental sessions were performed in eight rats (two or three sessions per rat). The impedance of the electrode-to-vagus nerve interface showed normal values in all rats during all experiments. The mean MST per rat during the baseline and VNS condition per session are shown in Table 1.

For each rat, one mean baseline MST and one mean VNS MST was calculated based on the different sessions (Fig. 1). Of the eight rats tested, seven showed an increase in MST with a delta value ranging between 258 and 784 μ A. One rat did not respond to the treatment.

During the VNS condition, the mean overall MST was significantly increased (1420 μ A, SEM 94 μ A) compared with the mean overall MST during the baseline condition (1072 μ A, SEM 54 μ A) (P < 0.01) (Fig. 2).

Also, during each individual cortical stimulation session a statistically significant increase in mean MST was found (P < 0.05, paired *t*-test) when baseline and VNS condition were compared. No significant differences were found between cortical stimulation sessions within the conditions. An overview of the data per session is shown in Table 2.

 $\label{eq:table_table} \begin{array}{c} \textbf{Table 1} & \text{Motor seizure threshold (MST) values during baseline and vagus nerve stimulation (VNS) conditions per rat per session \end{array}$

	Session	Mean MST baseline condition (SEM)	Mean MST VNS condition (SEM)
Rat 1	1	983 (189)	1524 (145)
	2	1461 (147)	1788 (59)
	3	1755 (51)	1829 (0)
Rat 2	1	1093 (91)	1687 (104)
	2	1227 (16)	1217 (27)
Rat 3	1	989 (132)	1999 (0)
	2	1001 (139)	1559 (124)
Rat 4	1	937 (32)	1413 (356)
	2	1413 (93)	1364 (97)
	3	968 (43)	1824 (121)
Rat 5	1	673 (25)	1037 (43)
	2	1084 (28)	1271 (28)
	3	1089 (63)	1597 (135)
Rat 6	1	908 (4)	957 (79)
	2	1323 (57)	1563 (57)
	3	876 (52)	1633 (96)
Rat 7	1	916 (55)	869 (44)
	2	983 (133)	1040 (53)
Rat 8	1	1112 (177)	1319 (92)
	2	824 (51)	1156 (31)
	3	1015 (15)	1252 (61)

Mean MST values during baseline and VNS conditions per rat are shown for each session. The baseline condition and VNS condition MST values were calculated as the mean of at least three consecutive stimulation trains. All values are presented in μA . SEM: standard error of the mean.

In total, five sessions of SHAM stimulation were performed in two rats (rat 3 and rat 4). The mean MST before SHAM stimulation (684 μ A) showed no difference from the mean MST after SHAM stimulation (694 μ A) (P > 0.05, paired Student's *t*-test).

EEG recording was used to confirm the focality of the induced epileptic seizures. During a focal seizure, no epileptiform discharges were observed



Figure 1. Motor seizure threshold (MST) values during baseline and vagus nerve stimulation (VNS) condition per rat. Mean MST values \pm 1 SEM during baseline and VNS condition per rat are shown. Values were calculated based on the different experimental sessions. Rat 2, 3 and 7 underwent two experimental sessions, the other five rats underwent three experimental sessions.



Figure 2. Overall motor seizure threshold (MST) value during baseline and vagus nerve stimulation (VNS) condition. The mean MST value for all rats during baseline (n = 8) and VNS (n = 8) condition is shown. *Statistically significant difference compared to the mean baseline MST value (paired Student's *t*-test; P < 0.05).

 Table 2
 Motor seizure threshold (MST) values during baseline and vagus nerve stimulation (VNS) conditions over consecutive experimental sessions

	Mean MST baseline condition (SEM)	Mean MST VNS condition (SEM)
Session 1 $(n = 8)$ Session 2 $(n = 8)$	951 (48) 1165 (80)	1351 (137)* 1370 (88)*
Session 3 $(n = 5)$	1141 (124)	1627 (83)*

Mean MST values during baseline and VNS conditions are shown for the number of rats tested in each session. *All sessions showed statistical significant increase in mean MST (P < 0.05) based on Student's *t*-test. All values are presented in μ A. SEM: standard error of the mean.

in the parietal locations posterior from the location of cortical stimulation, pointing to isolated epileptiform activity in the motor cortex, in agreement with the results of Krupp and Loscher (7). When stimulation was interrupted, the clinical seizure symptoms immediately stopped in all rats and no afterdischarges were identified on the EEG, confirming localized seizure activity restricted to a pure motor seizure.

Discussion

The data obtained in the present study demonstrate that VNS increases the threshold for focal motor seizures in the cortical stimulation rat model. It supports the hypothesis that VNS is able to modulate cortical excitability. These findings are in agreement with the reported acute antiseizure effect of VNS, that has been demonstrated in animal experiments and observed in clinical practice e.g. in patients who use the magnet feature of the device (8–12).

Our findings are in agreement with several other studies that have indicated a direct or indirect effect on cortical excitability of VNS. EEG studies in humans have shown an acute decrease in interictal epileptiform discharges after VNS, indicating a change in cortical neuronal activity (13, 14). Naritoku et al. found an increased latency in thalamocortical somatosensory evoked potentials after 1 month of VNS, suggesting modulation of the thalamocortical pathway by VNS (15). The nucleus of the solitary tract is the main terminal for vagal afferents in the brainstem and has direct and indirect projections to the locus coeruleus, the raphe nuclei, the reticular formation, the thalamus and ultimately the cortical neurons (1, 16). The anti-seizure effects of VNS could therefore be mediated through modulation of synaptic activity in the thalamus and the thalamocortical projection pathways. Several studies have shown VNSinduced changes in brain activation and cerebral blood flow in particular in the thalamus (17–20). Moreover, a positive correlation between thalamic activation and a favourable clinical outcome has previously been reported (17).

Based on *in vivo* intracellular cortical recordings, Zagon et al. proposed that slow hyperpolarization may be one of the mechanisms underlying the seizure-reducing effect of VNS, by means of reducing the excitability of neurons involved in seizure propagation (21). The acute effect of VNS on cortical excitability has also been investigated using transcranial magnetic stimulation (TMS). VNS was responsible for a pronounced increase in the inhibitory response produced by paired-pulse TMS but did not affect the excitatory response by single-pulse TMS. This observation specifically points to a GABA_A intracortical inhibitory mechanism explaining the modulatory effect of VNS on cortical excitability (22). The latter is in agreement with other studies demonstrating that GABA plays a major role in the MOA of VNS. Cerebrospinal fluid studies have shown an increase in GABA levels during VNS (23). A SPECT study in humans before and after 1 year of VNS treatment resulted into normalization of GABAA receptor density in patients with a clear therapeutic response (24). More recently, Neese et al. found that VNS following experimental brain injury in rats protects cortical GABAergic cells from cell death (25). Apart from the GABA hypotheses, there is experimental evidence indicating that also noradrenalin plays a role in the anti-seizure effect of VNS. Already in 1998, Krahl et al. demonstrated that lesioning the locus coeruleus, i.e. the major source of noradrenalin in the brain, abolished the seizureattenuating effect of VNS (26). A few years later, Groves et al. found a significant increase in the discharge rate of locus coeruleus neurons following short term VNS in the anaesthetized rat (27). Other experimental work confirmed this potential involvement of noradrenalin in the mechanism of action of VNS (28, 29). A pilot trial in our laboratory has shown responder-correlated increases in noradrenalin in the pilocarpine model using microdialysis (30).

This study shows efficacy of VNS in the cortical stimulation rat model using standard VNS stimulation parameters. These standard stimulation parameters are currently used in clinical practice, but are not evidence-based (2, 31, 32). This animal model may be a useful tool for future evaluation of optimized stimulation parameters as it may also be combined with microdialysis in freely moving animals. These studies may lead to improved clinical efficacy in patients treated with VNS.

In the present study, one rat did not show a significant increase in MST after VNS. We could

not identify any external factor causing the nonresponsiveness of this rat. However, a minor defect of the VNS electrode can not be excluded, even with normal impedance values for the electrodeto-vagus nerve interface. On the other hand, it is plausible that this finding reflects the non-responders also observed in clinical practice.

In this study VNS significantly increased the threshold for focal motor seizures in a cortical stimulation model. The obtained data indicate that VNS is capable of modulating electrically induced cortical excitability. Further research is needed to elucidate the precise mechanism of action of these VNS effects.

Acknowledgements

The authors wish to acknowledge Jeroen Van Aken for designing and programming the cortical stimulation pulse. Dr V. De Herdt is supported by junior researcher ('Aspirant') grant from the Fund for Scientific Research-Flanders (FWO). Prof. P. Boon is a Senior Clinical Investigator of the Fund for Scientific Research-Flanders and is supported by grants from FWO; grants from BOF and by the Clinical Epilepsy Grant from Ghent University Hospital. Prof. K. Vonck is supported by a BOF-ZAP grant from Ghent University Hospital. The VNS pulse generators used for these experiments were provided by Cyberonics Europe.

References

- HENRY TR. Therapeutic mechanisms of vagus nerve stimulation. Neurology 2002;59:S3–14.
- BOON P, DE HERDT V, VONCK K, VAN ROOST D. Clinical experience with vagus nerve stimulation and deep brain stimulation in epilepsy. Acta Neurochir Suppl 2007;97: 273–80.
- VOSKUYL RA, DINGEMANSE J, DANHOF M. Determination of the threshold for convulsions by direct cortical stimulation. Epilepsy Res 1989;3:120–9.
- DELLA PASCHOA OE, HOOGERKAMP A, EDELBROEK PM, VOSKUYL RA, DANHOF M. Pharmacokinetic-pharmacodynamic correlation of lamotrigine, flunarizine, loreclezole, CGP40116 and CGP39551 in the cortical stimulation model. Epilepsy Res 2000;40:41–52.
- HOOGERKAMP A, VIS PW, DANHOF M, VOSKUYL RA. Characterization of the pharmacodynamics of several antiepileptic drugs in a direct cortical stimulation model of anticonvulsant effect in the rat. J Pharmacol Exp Ther 1994; 269:521–8.
- JOSEPH S, DAVID J, JOSEPH T. Determination of anticonvulsant effects on seizure thresholds using ramp generated cortical stimulation in conscious rats. Indian J Exp Biol 1997;35:933–40.
- KRUPP E, LOSCHER W. Anticonvulsant drug effects in the direct cortical ramp-stimulation model in rats: comparison with conventional seizure models. J Pharmacol Exp Ther 1998;285:1137–49.
- ZABARA J. Inhibition of experimental seizures in canines by repetitive vagal stimulation. Epilepsia 1992;33:1005–12.
- BOON P, VONCK K, VAN WALLEGHEM P et al. Programmed and magnet-induced vagus nerve stimulation for refractory epilepsy. J Clin Neurophysiol 2001;18:402–7.

- DE HERDT V, WATERSCHOOT L, VONCK K et al. Vagus nerve stimulation for refractory status epilepticus. Eur J Paediatr Neurol 2009;13:286–9.
- WOODBURY JW, WOODBURY DM. Vagal stimulation reduces the severity of maximal electroshock seizures in intact rats: use of a cuff electrode for stimulating and recording. Pacing Clin Electrophysiol 1991;14:94–107.
- McLachlan RS. Suppression of interictal spikes and seizures by stimulation of the vagus nerve. Epilepsia 1993; 34:918–23.
- KUBA R, GUZANINOVA M, BRAZDIL M, NOVAK Z, CHRASTINA J, REKTOR I. Effect of vagal nerve stimulation on interictal epileptiform discharges: a scalp EEG study. Epilepsia 2002; 43:1181–8.
- SANTIAGO-RODRIGUEZ E, ALONSO-VANEGAS M, CARDENAS-MORALES L, HARMONY T, BERNARDINO M, FERNANDEZ-BOUZAS A. Effects of two different cycles of vagus nerve stimulation on interictal epileptiform discharges. Seizure 2006;15: 615–20.
- NARITOKU DK, MORALES A, PENCEK TL, WINKLER D. Chronic vagus nerve stimulation increases the latency of the thalamocortical somatosensory evoked potential. Pacing Clin Electrophysiol 1992;15:1572–8.
- RICARDO JA, KOH ET. Anatomical evidence of direct projections from the nucleus of the solitary tract to the hypothalamus, amygdala, and other forebrain structures in the rat. Brain Res 1978;153:1–26.
- 17. HENRY TR, VOTAW JR, PENNELL PB et al. Acute blood flow changes and efficacy of vagus nerve stimulation in partial epilepsy. Neurology 1999;**52**:1166–73.
- VONCK K, BOON P, VAN LAERE K et al. Acute single photon emission computed tomographic study of vagus nerve stimulation in refractory epilepsy. Epilepsia 2000;41:601–9.
- NARAYANAN JT, WATTS R, HADDAD N, LABAR DR, LI PM, FILIPPI CG. Cerebral activation during vagus nerve stimulation: a functional MR study. Epilepsia 2002;43:1509–14.
- LIU WC, MOSIER K, KALNIN AJ, MARKS D. BOLD fMRI activation induced by vagus nerve stimulation in seizure patients. J Neurol Neurosurg Psychiatry 2003;74:811–3.
- ZAGON A, KEMENY AA. Slow hyperpolarization in cortical neurons: a possible mechanism behind vagus nerve simulation therapy for refractory epilepsy? Epilepsia 2000;41: 1382–9.
- DI LAZZARO V, OLIVIERO A, PILATO F et al. Effects of vagus nerve stimulation on cortical excitability in epileptic patients. Neurology 2004;62:2310–2.
- 23. BEN-MENACHEM E, HAMBERGER A, HEDNER T et al. Effects of vagus nerve stimulation on amino acids and other metabolites in the CSF of patients with partial seizures. Epilepsy Res 1995;20:221–7.
- MARROSU F, SERRA A, MALECI A, PULIGHEDDU M, BIGGIO G, PIGA M. Correlation between GABA(A) receptor density and vagus nerve stimulation in individuals with drugresistant partial epilepsy. Epilepsy Res 2003;55:59–70.
- NEESE SL, SHERILL LK, TAN AA et al. Vagus nerve stimulation may protect GABAergic neurons following traumatic brain injury in rats: an immunocytochemical study. Brain Res 2007;1128:157–63.
- KRAHL SE, CLARK KB, SMITH DC, BROWNING RA. LOCUS coeruleus lesions suppress the seizure-attenuating effects of vagus nerve stimulation. Epilepsia 1998;39:709–14.
- GROVES DA, BOWMAN EM, BROWN VJ. Recordings from the rat locus coeruleus during acute vagal nerve stimulation in the anaesthetised rat. Neurosci Lett 2005;379:174–9.
- ROOSEVELT RW, SMITH DC, CLOUGH RW, JENSEN RA, BROWNING RA. Increased extracellular concentrations of norepinephrine in cortex and hippocampus following

vagus nerve stimulation in the rat. Brain Res 2006; 1119:124-32.

- 29. FOLLESA P, BIGGIO F, GORINI G et al. Vagus nerve stimulation increases norepinephrine concentration and the gene expression of BDNF and bFGF in the rat brain. Brain Res 2007;**1179**:28–34.
- 30. MEURS A, CLINCKERS R, RAEDT R et al. Vagus nerve stimulation suppresses pilocarpine-induced limbic seizures and

increases hippocampal extracellular noradrenalin concentration. Epilepsia 2008;**49**(S7):350.

- 31. BUNCH S, DEGIORGIO CM, KRAHL S et al. Vagus nerve stimulation for epilepsy: is output current correlated with acute response? Acta Neurol Scand 2007;**116**:217–20.
- LABINER DM, AHERN GL. Vagus nerve stimulation therapy in depression and epilepsy: therapeutic parameter settings. Acta Neurol Scand 2007;115:23–33.