"Transcriptomic biomarkers of human ageing in peripheral blood mononuclear cell total RNA"

Vo, Thi Kim Duy ; Godard, Patrice ; Morrhaye, Gabriel ; de Saint Hubert, Marie ; Bauwens, Emilie ; Debacq-Chainiaux, Florence ; Glupczynski, Gerald ; Swine, Christian ; Geenen, Vincent ; Martens, Henri J ; Toussaint, Olivier

ABSTRACT

Age-related changes of gene expression contribute to the physiological alteration observed with human ageing. Herein the abundance of a selection of 148 transcripts involved in immunosenescence and stress response was compared in total RNA of PBMC of healthy young to middle-age probands (35.0 +/- 6.5 year old) and healthy old probands (82.5 +/- 6.8 year old). This study provides a list of 16 differentially abundant transcripts species in the healthy old probands. Thus these changes of abundance can be considered as easily accessible biomarkers of ageing. Some of these differential abundances like CD28, CD69, LCK (decreased abundance in old subjects), CD86, cathepsin D, H and S (increased abundance in old subjects) might explain biochemical and cytochemical changes observed at the protein level in the immune system and thus might correspond to regulatory processes affecting the ageing process. Indeed these changes reflect the low-grade proinflammatory status observed in old persons and suggest a hypo-responsiveness of T-cells together with an increase in antigen presentation potential. In addition, among the differentially abundant transcripts were transcripts involved in the oxidative stress response HMOX1 and HSPA6 mRNAs were found as more abundant in PBMC from elderly subjects.

CITE THIS VERSION

Vo, Thi Kim Duy ; Godard, Patrice ; Morrhaye, Gabriel ; de Saint Hubert, Marie ; Bauwens, Emilie ; et. al. Transcriptomic biomarkers of human ageing in peripheral blood mononuclear cell total RNA. In: Experimental Gerontology, Vol. 45, no. 3, p. 188-194 (2010) http://hdl.handle.net/2078.2/28876 -- DOI : 10.1016/j.exger.2009.12.001
Transcriptomic biomarkers of human ageing in peripheral blood mononuclear cell total RNA

Thi Kim Duy Vo, Patrice Godard, Marie de Saint-Hubert, Gabriel Morrhay, Emilie Bauwens, Florence Debaq-Chainiaux, Youri Gluczynski, Christian Swine, Vincent Geenen, Henri J. Martens, Olivier Toussaint

1. Introduction

Ageing is defined as a progressive deterioration of biological functions (Faroqui and Faroqui, 2009) with progressive decreases of physiological capacity, reduced ability to cope with environmental stimuli, dysregulated inflammatory responses and changes in the immune response. Immunosenescence refers to the gradual age-related deterioration or modification of the immune system (Thoman and Weigle, 1989). Age-related changes of the immune system have been observed in both innate and acquired immunity (Huppert et al., 1998; Plackett et al., 2004). The effect of ageing on the functional capacity of T-cells was the most extensively studied. Some of the age-dependent biological changes that contribute to the onset of immunosenescence could result from the organism adaptation to the continuous challenge of adaptation to the environment. Ageing is accompanied by a chronic pro-inflammatory state, due to continuous antigenic stress that impinges upon innate immunity, throughout life, and has potential implications for the onset of inflammatory diseases (Vasto et al., 2007).

Transcriptomic profilings have been performed to study the effect of ageing on the regulation of gene expression in several cell types or organs (Galvin and Ginsberg, 2005; Vasto et al., 2007). Another purpose of gene expression profilings is to identify biomarkers that can be used later to better distinguish the state of physiological ageing of humans as opposed to their chronological ageing.

Herein, we developed a low-density DNA array to study the relative abundance of a selection of 148 transcripts species involved in immunosenescence, inflammation, stress response, cell or apoptosis regulation. The selection of 32 transcripts species involved in T-cell senescence was made upon survey of the literature. The selection of 116 transcripts species associated with stress response and cell senescence was also made according to literature survey. In addition, experimental work on cell senescence was also made according to literature survey. In several in vitro and in vivo experimental models (Borlon et al.,

Article history:
Received 16 September 2009
Received in revised form 25 November 2009
Accepted 1 December 2009
Available online 6 December 2009

Keywords:
Ageing
PBMC
Gene expression
Biomarker
Immunosenescence
Oxidative stress
cDNA array

Abbreviations: APC, antigen presenting cell; cDNA, complementary deoxyribonucleic acid; IFN-γ, interferon-γ; IL-10, interleukin-10; PBMC, peripheral blood mononuclear cells; RT-qPCR, retrotranscription quantitative polymerase chain reaction.

* Corresponding author. Tel.: +32 81 72 41 32; fax: +32 81 72 41 35. E-mail address: olivier.toussaint@fundp.ac.be (O. Toussaint).

2008a,b; de Magalhaes et al., 2004; Debacq-Chainiaux et al., 2005, 2008; Pascal et al., 2005; Zdanov et al., 2006). The reliability of the particular technology used herein was demonstrated previously in cell senescence-related studies and other biological models such as hepatic toxicity, adipocyte differentiation and oncology (de Lange et al., 2002; Gillet et al., 2004; Vankoningsloo et al., 2006). Inter- and intraplatform reproducibility of gene expression measurements was demonstrated with this technology (Patterson et al., 2006; Shi et al., 2006). Numerous verifications with real time retrotranscription quantitative polymerase chain reaction (RT-qPCR) have been done in all these models. For instance, in the emblematic model of replicative senescence of WI-38 human fibroblasts, comparison was also possible with many data previously found by other technologies like Northern blots or high density DNA arrays by many independent laboratories (Debacq-Chainiaux et al., 2008).

We report the identification of age-related transcriptomic biomarkers in total RNA of PBMC of old blood donors compared to young counter parts.

2. Materials and methods

2.1. Population recruitment

The recruitment was conducted at the University Hospital of Mont-Godinne (UCL, Belgium). We considered two groups: young to middle-age healthy volunteers and aged healthy volunteers. Young to middle-age participants were recruited on a voluntary basis among the staff of the hospital upon general announcement. The age of the young to middle-age volunteers ranged between 25 and 48 years old. All were clinically symptom-free. Healthy aged participants were recruited on a voluntary basis from different senior associations. To be eligible, participants needed (1) to be aged 75 years and over; (2) not to be institutionalized; (3) to have no evidence of an acute medical condition, nor deterioration of a chronic condition in the previous month. Exclusion criteria were the use of steroidal or non-steroidal anti-inflammatory drugs 1 week before the inclusion, cancer or cancer history, and haematological disease. The ethics committee of the hospital approved the study.

2.2. Isolation of PBMC and RNA expression

PBMCs were isolated from 16 mL of whole blood by centrifugation through Ficoll–Hypaque gradient centrifugation (Becton–Dickinson Vacutainer CPT, USA) and were washed twice in HBSS buffer. After adjustment at 10^6 cells/mL, part of the suspension were dispatched for hemogram determination (ADVIA 120, Baker, USA) while the rest were used for total RNA extraction (Total RNA-gents® Total RNA Isolation System, Promega, USA) according to the manufacturer's protocol and stored at −80 °C. The quality of total RNA samples was assessed (Agilent 2100 Bioanalyzer, Agilent Technologies, Germany). Reverse transcription, RNA amplification and labelling using the MessageAmp™ II-Biotin Enhanced kit (Ambion, USA) were performed from 250 ng of total RNA.

2.3. Low-density DNA-array analysis

2.3.1. Design of the array

We purchased a low-density DNA array on-demand able to detect 148 different transcripts species (Eppendorf, Germany) involved in immunosenescence, and more specifically focusing the T-cell compartment. Transcripts involved in stress response and cell senescence was also considered as described in the introduction.

The method is based on a system with two assays per glass slide with three identical sub-arrays per assay. The sequences of the RNA covalently linked to the glass slide were carefully chosen by sequence comparison. Checks were made to ensure that no cross-hybridization takes place. Several positive and negative hybridization controls plus detection controls were spotted on the array in order to control the reliability of the experimental data.

2.3.2. Hybridization and detection

Hybridization on the array was carried out as described by the manufacturer using 15 μg of complementary DNA (cDNA). The hybridization reaction was performed overnight at 60 °C in a Thermoblock for DualChip® Slides used with a Thermomixer comfort (Eppendorf, Germany).

Detection was performed using a Cy3-conjugated IgG anti-biotin (Jackson Immuno Research Laboratories, USA). Fluorescence of the hybridized arrays was scanned using the Packard ScanArray (PerkinElmer, USA) at a resolution of 10 μm. To maximize the dynamic range of detection, the same arrays were scanned at three photomultiplier (PMT) gains for quantifying high- and low-copy expressed genes. The scanned 16-bit images were imported into the ImaGene 4.1 software (BioDiscovery, USA). The fluorescence intensity of each DNA spot (median intensity of each pixel present within the spot) was calculated using local median background subtraction. Intensity values from high-PMT-gain pictures were used, except in the case of saturated spots. In the latter case, intensity values from intermediate PMT-gain pictures (or from low PMT-gain pictures in case of spots also saturated in the intermediate PMT-gain pictures) were used after scale correction. Only spots with median intensity after background subtraction at least two-fold higher than their local background were taken into account. The median of the three intensity values of the triplicate DNA spots was used in further steps.

2.4. Data normalization and inter-batch correction

All micro-arrays results were normalized against a single reference applying a local weighted regression (Smyth et al., 2003). The micro-array was provided by Eppendorf in three batches. The systematic intensity bias detected between the production batches was corrected successfully. Four samples were hybridized four times onto arrays of the three different batches. For each spotted sequence, a correction factor between the first batch and each of the others was computed using these four samples. This spot and batch dependent correction factor was applied to normalize intensities of each array. These normalized and corrected intensities were used in further statistical analyses.

2.5. RT-qPCR

First-strand cDNA was synthesized from 1 μg of total RNA, in a 20 μL reaction mix, using 200 U Superscript II reverse transcriptase in the presence of 1 μg of oligo(dT) (Invitrogen, UK). The expression of five genes was assessed using qPCR (sequence of the primers: Table 1). Amplification reaction assays were performed in 25 μL total reaction containing 12.5 μL of Sybr Green PCR Mastermix, 2.5 μL of each primer at 300 mM final concentrations and 5 μL of 1000-dilution of cDNA. qPCRs without cDNA were performed as template-free negative controls. qPCR were performed in duplicates with a hot start at 95 °C for 5 min followed by 40 cycles at 95 °C for 15 s and 65 °C for 1 min using the 7000 SDS thermal cycler (Applied Biosystems, The Netherlands). Melting curves were generated after amplification. Fluorescence emission was
detected for each PCR cycle and the threshold cycle values \( C_t \) were determined using the thermal cycler software. For each experiment and each gene the efficiency of the qPCR was assessed with three dilution factors (100; 1000 and 10,000) of one cDNA sample. The \( RPS9 \) gene was used as a housekeeping gene in order to compare gene expression between samples. This gene is selected for its expression constant in all the tested samples. The expression ratio of one gene \( g \) between two samples A and B was computed as following: 

\[
\frac{E_g^A}{E_g^B} \times \frac{E_r^A}{E_r^B},
\]

where \( E_g \) and \( E_r \) are efficiencies of qPCR for genes \( g \) and \( RPS9 \), \( C_{g,A} \), \( C_{g,B} \), \( C_{r,A} \) and \( C_{r,B} \) are the \( C_t \) of genes \( g \) and \( RPS9 \) in samples A and B.

2.6. Statistical analyses

The Mann–Whitney test was used to compare the difference of mRNA abundances between two groups. We used \( p \)-value with the Benjamini correction (\( p \)-value \( = \) m \( \times \) \( \alpha \), where \( m \) is the number of variables and \( g \) the rank of the variable according to \( p \)-value). The correlation between micro-array and RT-qPCR results was performed using a Pearson correlation analysis.

3. Results

We designed a specific low-density DNA array for studying mRNA abundance of gene involved in T-cell senescence, cell senescence in general and stress response. We explained in the introduction how this list of transcripts was designed. This list is available as Supplementary material (Table S1).

3.1. Mean age of the probands

RNA was isolated from PBMC of 25 young to middle-age healthy volunteers (14 women and 11 men, 25–48 years old, mean age 35.0 ± 6.5; mean ± SD), and 23 aged healthy volunteers (12 women and 11 men, 75–103 years old, mean age 82.5 ± 6.8; mean ± SD) subjects. The two groups were matched for gender. The health status of the probands was determined as explained in Section 2.

3.2. Micro-array result validation

More than 100 verifications of data obtained in bioregontological studies with a similar array have already been done using RT-qPCR. However, the results obtained herein on a random selection of five transcripts among our list of transcripts were used to assess the quality of micro-assay measurements using RT-qPCR. Fourteen independent samples from blood of different persons were analysed both with the array and with RT-qPCR. The comparison between RT-qPCR cycle threshold normalized and micro-array log2 ratio value using a Pearson correlation analysis yielded a \( p \)-value of 0.001. The coefficient correlation was 0.94, indicating that the micro-array results were significantly correlated with the RT-qPCR values (Fig. 1). The amplitudes of the expression ratio logarithms resulting from the RT-qPCR are generally slightly higher, as reported previously. The difference can be explained by a greater sensitivity of the RT-qPCR due to exponential amplification of the signal (Borlon et al., 2008a,b; Debacq-Chainiaux et al., 2005, 2008).

3.3. Identification of differentially abundant transcripts

From a total of 148 transcript species including those of 13 housekeeping genes, 77 transcript species were detected in the samples of PBMC of all the subjects, 71 transcript species were either not detectable in any of the subjects or not quantifiable. Thus 77 detected transcript species were selected as eligible for further analyses. There was no situation where a transcript was detected in a condition and not detected in the other.

We did no observe any significant differences of the abundance of each transcript between men and women in each age group of probands. To elucidate differences of the abundance of PBMC transcripts between young to middle-age and aged healthy participants, mRNA level was analysed individually for the 77 detected transcripts using a Mann–Whitney test, with a Benjamini and Hochberg false-discovery rate, and a \( p \)-value \( \leq 0.05 \). This resulted in a list of 16 differentially abundant transcripts. From these 16 transcripts, 11 were more abundant and five were less abundant in the PBMC of the elderly subjects. The list of these transcripts and a heat map are shown in Table 2 and Fig. 2, respectively.

3.4. Age-associated differential expression of genes in PBMC

The transcripts differentially abundant in PBMC of aged versus young to middle-age subjects, might be potential markers of
healthy ageing. Most of these transcripts differentially expressed in aged and young to middle-age groups can be sorted according to their variation with ageing. The transcripts less abundant with age were: CD28, CD69, LCK, PRDX6 and MAX. Transcripts more abundant in the aged group included CD86, CTSD, CTSH, CTSS, EIF4A1, GAA, HMox1, HSPA6, TNFRSF1A, TNFRSF1B and TUBA1B. The most important variations observed were increased abundance of HSPA6 and TNFRSF1A, respectively, 1.83- and 1.68-fold, and the decreased abundance of CD28 and CD69, respectively, 1.61- and 1.43-fold, in the aged group compared to the young group (Table 2 and Fig. 2). However, we have currently no available data to evaluate the abundance of the corresponding proteins.

### 4. Discussion

Ageing is associated with a myriad of changes in all the organs of our body. Ageing is a continuous process that affects individuals from maturity to old age, first gradually from 20 to 65 years old in humans. Abrupt changes are observed in healthy individuals over 75 years old (Faria et al., 2008).

In this work, we searched for transcriptional markers of human ageing in the total RNA of PBMC isolated from young to middle-age healthy (25–48 years old) and aged healthy (75–103 years old) probands. The study design was to select a limited panel of transcript species and to measure their abundance in PBMC samples from subjects expected to be in a sufficient number to reach statistical significances. The selected panel contained transcripts involved in the cellular mechanism of senescence, stress response as well as specific transcripts previously proposed to be involved in immunosenescence (Franceschi et al., 1999; Pawelec et al., 1999).

The comparison of data from young to middle-age and old probands showed a similar transcript abundance for about 79% of the transcripts while 21% displayed an altered abundant level in the aged group compared to the young group (Table 2 and Fig. 2). Statistical analysis was carried out with the Mann–Whitney test with a Benjamini and Hochberg false discovery threshold on 0.05. Fold change is calculated from the mean log ratio of aged versus young to middle-age.

Table 2

<table>
<thead>
<tr>
<th>Official gene symbol</th>
<th>Gene name (according to HUGO database and Entrez gene)</th>
<th>Accession number</th>
<th>Aged/young to middle-age</th>
<th>p-Value</th>
<th>Benjamini</th>
</tr>
</thead>
<tbody>
<tr>
<td>HSPA6</td>
<td>Heat shock 70-kDa protein 6 (HSP70B')</td>
<td>NM_002155.1</td>
<td>1.83</td>
<td>1.62 E–04</td>
<td>1.59 E–03</td>
</tr>
<tr>
<td>TNFRSF1A</td>
<td>Tumor necrosis factor receptor superfamily, member 1A</td>
<td>NM_001065.2</td>
<td>1.68</td>
<td>3.17 E–05</td>
<td>3.74 E–04</td>
</tr>
<tr>
<td>HMox1</td>
<td>Heme oxygenase (decycling) 1</td>
<td>NM_002133.1</td>
<td>1.46</td>
<td>1.62 E–05</td>
<td>3.19 E–04</td>
</tr>
<tr>
<td>GAA</td>
<td>Glucosidase, alpha; acid (Pompe disease, glycogen storage disease type II)</td>
<td>NM_000152.2</td>
<td>1.45</td>
<td>3.10 E–04</td>
<td>2.49 E–03</td>
</tr>
<tr>
<td>CTSH</td>
<td>Cathepsin H</td>
<td>NM_0004390.1</td>
<td>1.38</td>
<td>2.84 E–05</td>
<td>3.87 E–04</td>
</tr>
<tr>
<td>CTSD</td>
<td>Cathepsin D (lysosomal aspartyl peptidase)</td>
<td>M11233.1</td>
<td>1.37</td>
<td>3.38 E–06</td>
<td>1.20 E–04</td>
</tr>
<tr>
<td>CTSS</td>
<td>Cathepsin S</td>
<td>NM_006961.1</td>
<td>1.34</td>
<td>3.35 E–08</td>
<td>1.48 E–06</td>
</tr>
<tr>
<td>TNFRSF1B</td>
<td>Tumor necrosis factor receptor superfamily, member 1B</td>
<td>NM_001066.1</td>
<td>1.26</td>
<td>7.41 E–03</td>
<td>3.54 E–02</td>
</tr>
<tr>
<td>TUBA1B</td>
<td>Tubulin, alpha 1b</td>
<td>NM_006082</td>
<td>1.23</td>
<td>3.84 E–06</td>
<td>1.13 E–04</td>
</tr>
<tr>
<td>EIF4A1</td>
<td>Eukaryotic translation initiation factor 4A, isoform 1</td>
<td>NM_001416.1</td>
<td>1.22</td>
<td>6.31 E–06</td>
<td>1.60 E–04</td>
</tr>
<tr>
<td>CD86</td>
<td>CD86 antigen (DC28 antigen ligand 2, B7-2 antigen)</td>
<td>NM_006889.1</td>
<td>1.22</td>
<td>9.59 E–03</td>
<td>4.14 E–02</td>
</tr>
<tr>
<td>MAX</td>
<td>MHC associated factor X</td>
<td>NM_002382.1</td>
<td>0.78</td>
<td>2.04 E–05</td>
<td>3.00 E–04</td>
</tr>
<tr>
<td>PRDX6</td>
<td>Peroxiredoxin 6</td>
<td>NM_004905.1</td>
<td>0.77</td>
<td>1.82 E–05</td>
<td>3.22 E–04</td>
</tr>
<tr>
<td>LCK</td>
<td>Lymphocyte-specific protein tyrosine kinase</td>
<td>NM_005356.2</td>
<td>0.77</td>
<td>4.43 E–04</td>
<td>3.41 E–03</td>
</tr>
<tr>
<td>CD28</td>
<td>CD28 antigen (Tp44)</td>
<td>NM_006139.1</td>
<td>0.62</td>
<td>1.93 E–03</td>
<td>1.14 E–02</td>
</tr>
<tr>
<td>CD69</td>
<td>CD69 antigen (p60, early T-cell activation antigen)</td>
<td>NM_001781.1</td>
<td>0.70</td>
<td>7.41 E–04</td>
<td>3.75 E–02</td>
</tr>
</tbody>
</table>

Fig. 2. Clustered heat map of the most significantly differentially expressed genes in the PBMC between aged healthy and young to middle-age healthy probands. Differences in the levels of mRNA abundance are represented by intensity of color with red representing higher and green for lower abundance. The expression profiles show a significant difference between the aged healthy and the young to middle-age healthy groups. Gene symbols for each probe are listed under "gene", p-values are from a Mann–Whitney test with a Benjamini and Hochberg false discovery threshold on 0.05. Fold change is calculated from the mean log ratio of aged versus young to middle-age.
membrane glycoprotein, which plays multiple roles such as amplifying the cytokine production or insuring a complete and efficient T-cell activation (Nel, 2002; Riley and June, 2005). TCR engagement recruits LCK and FYN, protein kinases initiating the TCR signaling pathway (Salmond et al., 2009). Unlike LCK, expression of FYN was not affected by ageing in our study (data not shown). Decreased abundance of CD28 and LCK mRNA indicates that early signaling events following antigen stimulation are altered with ageing in T lymphocytes. This hypothesis of lowered activation was reinforced by the 30% decrease of the mRNA level of early activation surface markers CD69 found in comparison to young to middle-age subjects (Table 2 and Fig. 2). The changes at the mRNA level positively correlate with biochemical and cytometric observations (Boucher et al., 1998; Fulop et al., 1999; Lio et al., 1996). These age-related variations in the T-cell activation mechanisms probably explain the lower efficiency of vaccination procedure in old persons, where TCR antigen recognition and T-cell activation are primary phenomena involved as well as higher infectious risk when involving adaptive immune system.

Interestingly, the ligand of CD28 is a heterodimer formed by CD80 and CD86 present on many cell types with an antigen presenting cell (APC) activity (Lindsten et al., 1993; Salomon and Bluestone, 2001; Thompson et al., 1989). Up-regulation of CD80 in older adult monocytes by Toll-like receptor signaling was recently shown to be less effective while more complex regulation of CD86, which involves a secondary down-regulation by interleukin-10 (IL-10), also seems to be impaired in old subjects (van Duin et al., 2007). Nevertheless, a closer look at the dendritic cells from aged subjects revealed that both their CD80 and CD86 can be more efficiently up-regulated by self-antigen than in young subjects (Agrawal et al., 2009). We observed that, in healthy elderly, the basal level of expression of CD86 was 22% higher than in the young to middle-age group with no significant change in CD80. This higher level of CD86 could be related to the pro-inflammatory cytokine profile observed in the elderly subjects (Fagiolo et al., 1993). The CD86 down regulates IL-10 (Lim et al., 2002) not was evidenced significantly differences between healthy young and old probands (not shown). This suggests that the age-related potent increased co-stimulatory activity of APC is due to the elevated concentration of pro-inflammatory cytokines, which is counterbalanced by a reduced expression of CD28 receptor that could in the end lead to hypo-responsiveness of T-cell response.

Further involvement of ageing in the mechanism of antigen presentation is strongly suggested by a series of lysosomal proteases, which mRNA is more abundant in the elderly group. The Cathepsins genes CTSD, CTSH and CTSS all code for lysosomal proteases involved in physiological protein degradation (Kirschke and Wiedanders, 1987). Cathepsins are constitutively expressed in all known cell types. HMOX1 gene encodes heme oxygenase, which is an essential antioxidant enzyme.

Expression of GAA, EIF4A, MAX and TUBA1B is altered in ageing with increased mRNA abundance of GAA, EIF4A and TUBA1B transcripts and decreased abundance of MAX transcripts. The GAA gene encodes acid alpha-glucosidase, which is essential for the degradation of glycogen to glucose in lysosomes. The EIF4A1 gene is implied in the control of translation, and thus is involved in protein synthesis and cell proliferation (Flynn and Proud, 1996). The MAX gene encodes a protein of transcription regulator. The TUBA1B gene encodes the tubulin, which is the major constituent of microtubules. These are potential interesting transcript markers in ageing.

Ageing is associated with progressively increased oxidative stress and damage, with potential changes of expression of genes involved in stress response. In this work, HMOX1 and HSPA6 mRNAs were found as more abundant in PBMC from elderly subjects, compared with PBMC from young to middle-age control subjects. HMOX1 gene encodes heme oxygenase, which is an essential enzyme associated with oxidative stress in many cell types such as fibroblasts (Pascal et al., 2007). HSPA6 is a member of the human heat-shock protein gene family, encoding a basic 70-kDa protein (Leung et al., 1990). HMOX1 and HSPA6 mRNA levels are potential functional markers of ageing, probably due to their involvement in stress response (Calabrese et al., 2008; Chowdhury et al., 2009; Gong et al., 2004; Hori et al., 2002; Lee et al., 1996). HMOX1 activity and protein levels were increased in the liver with age (Abraham et al., 1985; Kang et al., 2005). As for HSPA6, it was reported that aged individuals of particular species develop a constitutively increased level of several chaperones, including HSP70 (Soti and Csermely, 2000). The finding that HMOX1 and HSPA6 transcript species increased in healthy old persons may be explained by compensatory induction of these genes to cope with increased oxidative stress and damage in ageing. We have no clue about the potential reasons for the decreased mRNA level of PRDX6 antioxidant enzyme.

Ageing is a multi-dimensional process. Age-related changes of gene expression contribute to the physiological alterations observed with ageing. This work provided a list of 16 transcripts dif-

The plausible consequences of increased pro-inflammatory cytokine profile of old persons are known. In this study, we observed an up-regulation of some of these cytokine receptors at the mRNA level. The TNFRSF1A and TNFRSF1B genes encode the type 1 (TNFR-1), and type 2 (TNFR-2) receptors of TNF-α, respectively, and exert their biological through distinct signaling pathways (Gupta et al., 2004). In addition to inflammatory cytokine induction, TNF-α can stimulate apoptosis, but also activates NF-κB transcription factor, which in turn activates transcription of genes that block TNF-induced cell death (Xiao et al., 2001, 2002). With ageing, both TNF-α production and TNF-α-induced apoptosis are increased (Agarwal et al., 1999; Fagiolo et al., 1993). An increased expression of TNFRSF1A and a decreased expression of TNFRSF1B in lymphocytes from aged humans have been reported (Agarwal et al., 1999). These changes have been associated with increased apoptosis observed in aged lymphocytes, as it was suggested that TNFRSF1A pathway is more pro-apoptotic while TNFRSF1B pathway would preferentially induce the anti-apoptotic c-IAP1 and c-IAP2 proteins. In our study, however, both TNFRSF1A and TNFRSF1B are up-regulated in PBMC from aged humans. Our results, if confirmed at protein level by further studies, would be in contradiction with the hypothesis of an overall pro-apoptotic mechanism in aged T-cell and suggests that a balance might emerge between the two receptors, leading to a more subtle regulation of T-cell apoptosis.

Aside from the modifications observed in the abundance of transcripts directly involved in the immune system function, other transcripts displayed significantly modified abundance in the PBMC of the old probands.

Expression of GAA, EIF4A, MAX and TUBA1B is altered in ageing with increased mRNA abundance of GAA, EIF4A and TUBA1B transcripts and decreased abundance of MAX transcripts. The GAA gene encodes acid alpha-glucosidase, which is essential for the degradation of glycogen to glucose in lysosomes. The EIF4A1 gene is implied in the control of translation, and thus is involved in protein synthesis and cell proliferation (Flynn and Proud, 1996). The MAX gene encodes a protein of transcription regulator. The TUBA1B gene encodes the tubulin, which is the major constituent of microtubules. These are potential interesting transcript markers in ageing.

Ageing is associated with progressively increased oxidative stress and damage, with potential changes of expression of genes involved in stress response. In this work, HMOX1 and HSPA6 mRNAs were found as more abundant in PBMC from elderly subjects, compared with PBMC from young to middle-age control subjects. HMOX1 gene encodes heme oxygenase, which is an essential enzyme associated with oxidative stress in many cell types such as fibroblasts (Pascal et al., 2007). HSPA6 is a member of the human heat-shock protein gene family, encoding a basic 70-kDa protein (Leung et al., 1990). HMOX1 and HSPA6 mRNA levels are potential functional markers of ageing, probably due to their involvement in stress response (Calabrese et al., 2008; Chowdhury et al., 2009; Gong et al., 2004; Hori et al., 2002; Lee et al., 1996). HMOX1 activity and protein levels were increased in the liver with age (Abraham et al., 1985; Kang et al., 2005). As for HSPA6, it was reported that aged individuals of particular species develop a constitutively increased level of several chaperones, including HSP70 (Soti and Csermely, 2000). The finding that HMOX1 and HSPA6 transcript species increased in healthy old persons may be explained by compensatory induction of these genes to cope with increased oxidative stress and damage in ageing. We have no clue about the potential reasons for the decreased mRNA level of PRDX6 antioxidant enzyme.

Ageing is a multi-dimensional process. Age-related changes of gene expression contribute to the physiological alterations observed with ageing. This work provided a list of 16 transcripts dif-
ferentially abundant in the PBMC of old healthy subjects, compared to subjects of young to middle-age. Even with the selection bias of gene selection and PBMC examination, changes of expression of immune response-related genes appears predominant in this study. Most of these changes can be related to modification of immune functions observed in aged healthy people. Thus, the transcriptomic changes identified herein should not be regarded only as biomarkers since these alterations might also have physiological consequences.

Acknowledgements

The authors thank all the patients included in this study for their participation. This work was supported by the Walloon Region of Belgium (DGTRE Reseaux 2-SENEGNE No. 05/1/6192). O. Toussaint and F. Debaq-Chainiaux are, respectively, Senior Research Associate and Research Associate of the FNRS Belgium.

Appendix A. Supplementary data


References


