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Protective Antioxidant and Antiapoptotic Effects of ZnCl₂ in Rat Pancreatic Islets Cultured in Low and High Glucose Concentrations

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Introduction

Type 2 diabetes results from the combination of insulin resistance and defective glucose stimulation of insulin secretion by the endocrine pancreas. The latter defect is due to a reduction in pancreatic β-cell mass and function [1,2] that has been diversely attributed to low grade inflammation, mitochondrial oxidative stress or endoplasmic reticulum stress [3]. In this context, we and others have previously shown that, after prolonged culture in the presence of a large range of glucose concentrations, rat islet cell apoptosis follows an asymmetric V-shaped profile with a minimum in 10 mmol/l glucose (G10), a large increase in 5 mmol/l glucose (G5) and moderately increased in 30 mmol/l glucose (G30). This glucose-dependent asymmetric V-shaped profile is preceded by parallel changes in the mRNA levels of oxidative stress-response genes like Metallothionein 1a (Mt1a). In this study, we tested the effect of ZnCl₂, a potent inducer of Mt1a, on apoptosis, mitochondrial oxidative stress and alterations of glucose-induced insulin secretion (GSIS) induced by prolonged exposure to low and high vs. intermediate glucose concentrations.

Methods: Male Wistar rat islets were cultured in RPMI medium. Islet gene mRNA levels were measured by RTq-PCR. Apoptosis was quantified by measuring islet cytosolic histone-associated DNA fragments and the percentage of TUNEL-positive β-cells. Mitochondrial thiol oxidation was measured in rat islet cell clusters expressing “redox sensitive GFP” targeted to the mitochondria (mt-roGFP1). Insulin secretion was measured by RIA.

Results: As observed for Mt1a mRNA levels, β-cell apoptosis and loss of GSIS, culture in either G5 or G30 vs. G10 significantly increased mt-roGFP1 oxidation. While TPEN decreased Mt1a/2a mRNA induction by G5, addition of 50–100 μM ZnCl₂ to the culture medium strongly increased Mt1a/2a mRNA and protein levels, reduced early mt-roGFP oxidation and significantly decreased late β-cell apoptosis after prolonged culture in G5 or G30 vs. G10. It did not, however, prevent the loss of GSIS under these culture conditions.

Conclusion: ZnCl₂ reduces mitochondrial oxidative stress and improves rat β-cell survival during culture in the presence of low and high vs. intermediate glucose concentrations without improving their acute GSIS.

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Conclusion: ZnCl₂ reduces mitochondrial oxidative stress and improves rat β-cell survival during culture in the presence of low and high vs. intermediate glucose concentrations without improving their acute GSIS.
Figure 1. Effects of glucose and Zn\(^{2+}\) chelation by TPEN on Mt1a, Mt2a, ZnT1 and ZnT8 mRNA expression in overnight cultured rat islets. Rat islets were precultured for 1 week in serum-free RPMI medium containing G10 and 5 g/l BSA. They were then cultured for 18 h in the...
buffering capacity [15]. In addition, the increase in [Zn$^{2+}$], has also been proposed to exert antioxidant and antiapoptotic effects through various mechanisms, e.g. caspase inhibition, xanthine oxidase inhibition, increased cytosolic superoxide dismutase activity and metallothionein overexpression [16].

$Zn^{2+}$ is an important cofactor for insulin biosynthesis and crystallization [17] and is co-secreted with insulin. A non-synonymous polymorphism in the $Slc30a8$ gene encoding the insulin granule $Zn^{2+}$ transporter $Slc30a8$ ($ZnT8$) has been associated with the risk of developing type 2 diabetes [18]. Interestingly, type 2 diabetic patients display a marked decrease in total plasma $Zn^{2+}$ concentration together with hyperzincuria, and $Zn^{2+}$ supplementation has been shown to ameliorate glycemic control in both type 1 and type 2 diabetes (reviewed in [19,20]. A high-$Zn^{2+}$ diet also improved blood glucose levels after islet transplantation in diabetic rats [21]. However, the mechanisms underlying these beneficial effects of $Zn^{2+}$ have been poorly elucidated.

We therefore tested the effect of $ZnCl_2$, a potent inducer of metallothionein expression, on islet cell apoptosis and the alterations of glucose-stimulated insulin secretion (GSIS) after prolonged culture of rat islets in low and high $Zn$ intermediate glucose concentrations. Using the mitochondria-targeted redox-sensitive ratiometric fluorescent probe mt-roGFP1, we also tested the effect of culture in the presence of low, intermediate and high glucose concentrations with or without $ZnCl_2$ on $\beta$-cell mitochondrial thiol oxidation state.

### Materials and Methods

#### Materials

Dithiothreitol (DTT) and N,N,N,N’-tetraakis(2-pyridyl-methyl)-ethylenediamine (TPEN) were purchased from Sigma (St-Louis, MO, USA). Hydrogen peroxide ($H_2O_2$) and $ZnCl_2$ were obtained from Acros Organics (Thermo Fisher Scientific, New Jersey, USA). Other reagents of analytical grade were purchased from Merck (Darmstadt, Germany).

#### Islet Isolation and Culture

Pancreatic islets were isolated from $\sim$200 g male Wistar rats as described [4]. Except for experiments on cell clusters, the islets were pre cultured for one week at 37°C and 5% $CO_2$ in serum-free RPMI 1640 (Invitrogen, Carlsbad, CA, USA) containing 5 g/l BSA and 10 mmol/l glucose. They were then cultured for up to 1 week in the same medium containing 5, 10 or 30 mmol/l glucose (G5, G10, or G30) and various test substances, and processed for further analysis. Experimental procedures were approved by the local ethics committee for animal experimentation.

#### Real-time PCR

Islet gene mRNA levels were measured as described [22]. Briefly, islet total RNA was extracted using Tripure (Roche Diagnostics GmbH, Mannheim, Germany) and reverse transcribed into cDNA using 50 ng of randoms hexamers and 200 units of the enzyme RevertAidTM H Minus M-MuLV Reverse Transcriptase (Fermentas GmbH, St.Leon-Rot, Germany). Real-time PCR was performed with an iCycler iQ Real-Time PCR Detection System (Bio-Rad, Hercules, CA). Primers sequences and reactions conditions are shown in Table S1. Islet gene mRNA to TATA-box binding protein ($Tbp$) mRNA or cyclophilin mRNA ratios ($2^{-\Delta\Delta Ct}$) were expressed relative to the ratio in islets cultured in G10.

#### Immunodetection of MT1a/2a

Islets were fixed in 4% paraformaldehyde and embedded in paraffin. Five $\mu$m thick sections were incubated overnight with a mouse anti- $MT1a/2a$ antibody (Abcam, Cambridge, UK) diluted 1:100, washed in Tris-buffered saline and incubated for 1 h with Alexa Fluor 488-conjugated goat anti-mouse IgG (Invitrogen, CA, USA) diluted 1:200. Sections were then mounted with Vectorshield-mounting medium containing 4’,6-diamidino-2-phenylindole (DAPI) (Vector Laboratories, Burlingame, CA) and visualized on a fluorescence microscope (FluoArc mounted on an Axioskop 40 microscope coupled to an HBO 100 camera; Carl Zeiss, Oberkochen, Germany) under standardized conditions (excitation/emission wavelengths: insulin: 475/540 nm; DAPI: 350/460 nm; $MT1a/2a$: 590/617 nm).

#### Islet Cell Apoptosis

Cytoplasmic histone-associated DNA fragments were measured on batches of 50 to 60 islets using the Cell Death Detection ELISA PLUS kit (Roche Diagnostics) as described [22]. The percentage of apoptotic $\beta$-cells (TUNEL-positive and insulin-positive cells) was determined on 5 $\mu$m thick islet sections using the In Situ Cell Death Detection Kit - POD (Roche Diagnostics) following the manufacturer’s instructions. Sections were then incubated overnight with a guinea-pig anti-insulin antibody (1/1000) (Invitrogen, Eugene, Oregon, USA) and incubated for 1 h with an Alexa Fluor 488-conjugated anti-guinea-pig antibody (1/1000) (Invitrogen, Caralillo, CA93012, USA). Islet cell nuclei were stained with DAPI. The percentage of apoptotic $\beta$-cells was determined by manually counting TUNEL, DAPI and insulin-positive nuclei on digital images obtained by fluorescence microscopy (FluoArc mounted on an Axioskop 40 coupled to a HBO 100 camera; Carl Zeiss, Oberkochen, Germany) under standardized conditions (excitation/emission wavelengths: fluorescein: 475/540 nm; DAPI: 350/460 nm; insulin: 390/617 nm).

#### Adenovirus

Adenovirus encoding mt-roGF1 under the control of the CMV promoter were generated and amplified using the pAdEasy system (Strategene, La Jolla, CA), as previously described [11]. After purification on a gradient of CsCl, the infectious titre of viral stocks was determined with the Adeno-X™ Rapid Titer kit (Clontech, Mountain View, CA, USA).

#### Mitochondrial Oxidative Stress Measurement

Mt-roGF1, which measures the thiol/disulfide equilibrium in the mitochondrial matrix, was used as an indicator of mitochondrial redox status [23,24]. After isolation, rat islets were dispersed...
in clusters using trypsin and gentle pipetting in a Ca²⁺-free medium. Cells clusters were plated on glass coverslips and cultured overnight in RPMI 1640 medium containing G10 and 10% Fetal Bovine Serum (FBS). Cells were infected for 48 h with adenovirus coding mt-roGFP1 (multiplicity of infection ~25 to 50) and the medium was changed for the last 10 to 24 h with a medium containing G5 or G10 and FBS with or without 50 μM ZnCl₂.

**Mt1a/Mt2a fluorescence** (excitation: 405/480 nm; emission: 535 nm, 40X objective) was measured every 30 s in cell clusters perfused with a bicarbonate-buffered Krebs solution containing (mmol/l) NaCl (120), KCl (4.8), CaCl₂ (2.5), MgCl₂ (1.2), NaHCO₃ (24), 1 g/l BSA (fraction V, Roche, Basel, Switzerland) and continuously gassed with O₂/CO₂ (94/6) to maintain pH ~7.4. For the first 20 min, cells were perfused with the same glucose concentration as during the last period of culture but without ZnCl₂. They were then perfused with 10 mmol/l DTT for 15 min to maximally reduce the probe, followed by 1 mmol/l H₂O₂ for 25 min to maximally oxidize mt-roGFP1. The ratio of fluorescence intensities (exc 405/480) were computed and expressed as a percentage of the difference between the mean ratio measured from 4 to 8 min after addition of DTT (set at 0%) and that measured from 14 to 18 min after addition of H₂O₂ (set at 100%).

**Insulin Secretion**

After culture, batches of 5 islets were incubated for 45 min in a bicarbonate-buffered Krebs solution containing 0.5 mmol/l glucose. Islets were then incubated for 1 h in the presence of various glucose concentrations. Insulin concentration in the medium was measured by RIA using rat insulin as a standard [25], and normalized for variations in islet DNA content measured by fluorimetry using SYBR Green I [26].

**Statistical Analysis**

Results are means ± SEM for the indicated number of experiments. Statistical significance of differences between groups was assessed by one-way ANOVA followed by a test of Newman-Keuls or by two-way ANOVA followed by a test of Bonferroni, as indicated in the legends. Differences were considered significant if P<0.05.

**Results**

**Effects of Glucose and Zn²⁺ Chelation on Mt1a, Mt2a, ZnT1 and ZnT8 mRNA Expression in Overnight Cultured Rat Islets**

After overnight culture of whole rat islets in the presence of 5, 10 or 30 mmol/l glucose (G5, G10 or G30), islet gene mRNA levels for the typical MTF-target genes Mt1a and Mt2a were minimal in G10, largely increased in G5, and slightly but not significantly increased in G30 (Fig. 1A-D). In comparison, the mRNA levels of the other MTF-target gene Slc30a1 (Znt1) only increased ~2-fold in G5 vs. G10 and were not affected by G30 (Fig. 1E,F). In contrast, Slc30a8 (Znt8) mRNA expression decreased after culture in either G5 or G30 vs. G10 (Fig. 1G,H). Under these conditions, the membrane-permeable Zn²⁺ chelator TPEN almost fully inhibited the stimulation of Mt1a, Mt2a mRNA expression by culture in G5, suggesting that a rise in [Zn²⁺], is involved in this effect of low glucose. In contrast, TPEN failed to affect Mt1a and Mt2a mRNA levels in G10 and G30 but significantly reduced Znt1 and Znt8 mRNA levels under all conditions. These effects of TPEN were, however, accompanied by a ~2- to 4-fold reduction in the mRNA levels of the housekeeping genes Tbp and cyclophilin under all culture conditions, and by a clear increase in islet cell apoptosis (not shown).

**Long-term Effects of Glucose and ZnCl₂ on Mt1a, Mt2a, ZnT1 and ZnT8 mRNA Expression in Cultured Rat Islets**

After one week of culture in G5, Mt1a mRNA expression increased ~10 fold in comparison with islets cultured in G10 (Fig. 2A). However, contrasting with data obtained after 18 h of culture, Mt1a mRNA expression did not increase and even tended to decrease after one week of culture in G30 vs. G10 (Fig. 2B). Similar results were obtained for Mt2a while Znt1 and Znt8 mRNA levels were not affected by one week of culture at different glucose concentrations (Fig. 2C-H). Under these conditions, addition of 100 μmol/l ZnCl₂ to the medium increased Mt1a, Mt2a and Znt1 but not Znt8 mRNA expression at all glucose concentrations, Mt1a and Mt2a mRNA reaching levels at least a 100-fold higher than in islets cultured in G10 (Fig. 2). In comparison, addition of 10 or 30 μmol/l ZnCl₂ to G5 only slightly increased Mt1a mRNA expression 1.9±0.4 and 2.2±0.6 times respectively (20±4 and 24±7 times the level in G10-cultured islets, n = 3). We therefore used ZnCl₂ at the concentration of 100 μmol/l in all subsequent experiments carried out with whole islets.

**Long-term Effects of Glucose and ZnCl₂ on Mt1a/2a Protein Levels in Cultured Rat Islets**

As shown in figure 3, Mt1a/2a protein levels were not detectably affected by one week of culture in G5 or G30 vs. G10. However, addition of 100 μM ZnCl₂ strongly increased Mt1a/2a protein levels specifically in β-cells at all glucose concentrations.

**Effects of ZnCl₂ on Rat Islet Cell Apoptosis Induced by Prolonged Culture in Low and High Glucose**

We next tested the effect of ZnCl₂ on islet cell apoptosis induced by prolonged culture in extreme glucose concentrations. As shown in figure 4AD, three days of culture in G5 instead of G10 induced a ~3.5-fold increase in cytoplasmic histone-associated DNA fragments and a ~4.3-fold increase in the percentage of TUNEL-positive β-cell nuclei. Addition of ZnCl₂ during culture significantly reduced islet cell DNA fragmentation by ~38% and the percentage of apoptotic β-cells by ~68%. After one week of culture in G5, DNA fragmentation increased ~30-fold while the percentage of apoptotic β-cells increased ~12 fold (Fig. 4BE), and addition of ZnCl₂ to the medium
significantly reduced these effects by 27% and 70% respectively. After one week of culture in G30, islet cytoplasmic DNA fragments only tended to increase ~1.5 fold, but the percentage of apoptotic β-cells was ~6 times higher than in islets cultured in G10 (Fig. 4CF). Under these conditions, ZnCl₂ reduced the stimulation of DNA fragmentation by ~90% and the increase in the proportion of TUNEL-positive β-cells by 65%. These results indicate that 100 μmol/l ZnCl₂ exerts a protective effect against rat β-cell apoptosis induced by chronic exposure to extreme glucose concentrations.

Effects of Glucose and ZnCl₂ on Mitochondrial Oxidative Stress in Rat Islet Cell Clusters

As ZnCl₂ has been proposed to improve cell resistance to oxidative stress, we next tested its effect on early mitochondrial oxidative stress induced by extreme glucose concentrations in rat islet cell clusters expressing mt-roGFP1. In these preparations, the concentration of ZnCl₂ had to be reduced to 50 μmol/l to avoid a strong increase in mt-roGFP1 oxidation in G10-cultured clusters (data not shown). As shown in figure 5BC, overnight culture in the presence of G5 or G30 instead of G10 significantly increased mt-roGFP1 oxidation in rat islet cell clusters, reflecting an increase in

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**Figure 3. Long term effects of glucose and ZnCl₂ on MT1a/2a protein levels in cultured rat islets.** After preculture, rat islets were cultured for 1 week in the presence of G5, G10 or G30 alone or with 100 μmol/l ZnCl₂. Islets were then fixed in paraformaldehyde solution (4%) and embedded in paraffin. Nuclei (DAPI), insulin and MT1a/2a were detected by immunohistochemistry in 5 μm-thick islets sections. Bar scale = 100 μm. A-D: islets cultured in G5; E-H: islets cultured in G5+100 μM ZnCl₂ (G5+ Zn); I-L: islets cultured in G10; M-P: islets cultured in G10+100 μM ZnCl₂ (G10+ Zn); Q-T: islets cultured in G30; U-X: islets cultured in G30+100 μM ZnCl₂ (G30+ Zn). A,E,I,M,O,U: DAPI staining; B,F,J,N,R,V: insulin staining; C,G,K,O,S,W: MT1a/ 2a staining; D,H,L,P,T,X: merge. Results are representative for 2 to 3 experiments. doi:10.1371/journal.pone.0046831.g003

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**Figure 4. Effects of glucose and ZnCl₂ on the stimulation of rat islet cell apoptosis by prolonged culture in low and high glucose concentrations.** Rat islets were cultured for 3 days (A, D) or 1 week (B, C, E, F) in the presence of G5, G10 or G30+ 100 μmol/l ZnCl₂ as indicated. A, B, C, DNA fragmentation was measured by an ELISA kit and expressed relative to the value measured in islets cultured in G10. D, E, F, the percentage of apoptotic β-cells was measured by TUNEL. Results are means ± SEM for 3–5 experiments. *, P<0.05 for the effect of G5 or G30 vs. G10 and #, P<0.05 for the effect of ZnCl₂ by two-way ANOVA followed by a test of Bonferroni (A, B, C) or by one-way ANOVA followed by a test of Newman-Keuls (D, E, F). doi:10.1371/journal.pone.0046831.g004
thiol (e.g. glutathione) oxidation in the mitochondrial matrix. Addition of ZnCl2 to the culture medium diminished by \(\sim 40\%\) the level of mt-roGFP1 oxidation in G5, and tended to reduce by \(\sim 23\%\) that in G30. These results suggest that ZnCl2 exerts some antioxidant effect in rat islet cell clusters exposed to low and high \(\beta\)-cell glucose concentrations.

**Effects of ZnCl2 on the Alterations of Glucose-induced Insulin Secretion after Prolonged Culture in Low and High Glucose**

We finally tested the effects of ZnCl2 during culture on islet insulin content and acute GSIS. As shown in figure 6A,B,E,F,I, three days of culture in the presence of G5 decreased the islet insulin content by \(\sim 75\%\), decreased the absolute rate of insulin secretion in G0.5 and G6, and profoundly inhibited its stimulation by G10 and G20. Addition of ZnCl2 to the culture medium did not affect the islet insulin content or GSIS of islets cultured in G5, but significantly reduced the acute insulin secretory response to G10 of islets cultured in G10. In comparison with culture in G5, one week culture in G30 only reduced the islet insulin content by \(\sim 30\%\) and differently affected the GSIS, with a slight increase in the rate of insulin secretion in the presence of G6, and a reduced stimulation of insulin secretion in response to G10 and G20 (Fig. 6C,D,G,H,I). Although ZnCl2 prevented the decrease in islet insulin content during culture in G30, it did not improve their GSIS. The increase in insulin content was not due to an increase in preproinsulin mRNA levels (Figure 6K,L). Thus, despite the beneficial effect of ZnCl2 on \(\beta\)-cell survival, ZnCl2 did not protect against the alterations of GSIS induced by culture in extreme glucose concentrations.

**Discussion**

We have previously shown that prolonged culture of rat islets in the presence of low or high \(\beta\)-cell intermediate glucose concentrations rapidly induces the expression of oxidative stress-response genes such as \(\text{Mt}1\alpha\) and \(\text{Hmox}1\), followed by later increase in islet cell apoptosis and marked reduction of GSIS [4]. We now provide further evidence that these changes are associated with early parallel changes in mitochondrial oxidative stress, and demonstrate that ZnCl2, which potently induces \(\text{Mt}1\alpha\) and \(\text{Mt}2\alpha\) expression, exerts a protective effect on mitochondrial thiol oxidation and subsequent \(\beta\)-cell apoptosis without improving GSIS.

In isolated rat islets cultured overnight in the presence of increasing glucose concentrations, the mRNA levels of the MT1-target genes \(\text{Mt}1\alpha\) and \(\text{Mt}2\alpha\) were minimal in G10, markedly increased in G5, and tended to increase in G30 vs. G10. These glucose effects were associated with parallel changes in mt-roGFP1 oxidation, a good indicator of thiol (mainly glutathione) oxidation in the mitochondrial matrix [11,27], suggesting the presence of mitochondrial oxidative stress after 18h culture in either low or high \(\beta\)-cell intermediate glucose concentrations. We have recently shown that mt-roGFP1 oxidation is acutely stimulated in rat islet cell clusters upon a reduction in glucose concentration from 10 to 2 mmol/l but that it is not increased upon glucose stimulation from 10 to 30 mmol/l glucose [11]. Together with the present study, these data indicate that the stimulation of oxidative stress induced by G30 is slower than that induced by G5, as is the case for the stimulation of \(\beta\)-cell apoptosis under these culture conditions [4]. However, in both G5 and G30, mt-roGFP1 oxidation occurred earlier than \(\beta\)-cell apoptosis, suggesting that the latter may result from mitochondrial oxidative stress. In that scenario, we postulate that the increase in \(\text{Mt}1\alpha\) and \(\text{Mt}2\alpha\) mRNA levels are sensitive indicators of this type of stress. Although we did not fully investigate the mechanism of \(\text{Mt}1\alpha\) and \(\text{Mt}2\alpha\) mRNA inductions, it is possible that oxidation of metallothioneins releases Mt-bound Zn\(^{2+}\), with consequent activation of metal transcription factor-1 (MTF-1) and increased expression of its target genes \(\text{Mt}1\alpha\), \(\text{Mt}2\alpha\) and \(\text{Znt}1\) [28,29]. In comparison, expression of the type 2 diabetes gene \(\text{Skl}30\alpha\beta\) that encodes the \(\beta\)-cell specific granular zinc transporter \(\text{ZnT}8\) was not regulated in parallel with \(\text{Znt}1\) or \(\text{Mt}\), in...
agreement with the observations that it is not induced by ZnSO4 [30] nor by ZnCl2 (the present study). Despite the large changes in Mt1a and Mt2a mRNA levels, MT1a/2a protein levels were not detectably increased by culture in low or high vs. intermediate glucose concentrations. This discordance between changes in Mt mRNA and protein levels could result from the low sensitivity of immunohistochemistry and, at least in low glucose, from a global decrease in protein translation in G5 vs. G10 [31].

It has recently been shown that the free cytosolic Zn2+ concentration ([Zn2+]c) in CD1 mouse islets decreases from 800 to 400 pmol/l after 24 h culture in 3 vs. 16.7 mmol/l glucose while Mtf-2 mRNA levels increase and Zip6-8 expression decreases under these conditions [32]. Although these results seem to argue against a role of a rise in [Zn2+]c in the stimulation of Mt gene expression by culture in low glucose, our hypothesis is strongly supported by the observation that the membrane-permeable Zn2+ chelator TPEN fully suppressed the induction of MTF-target gene expression by G5. Thus, we interpret the late decrease in [Zn2+]c measured by Bellomo et al. after 24 h culture in 3 vs. 16.7 mmol/l glucose as a possible consequence of the increase in metallothionein expression (although it was not detected by immunohistochemistry) and Zn2+-buffering capacity under these culture conditions. Alternatively, glucose-induced changes in [Zn2+]c might be different in the cytosolic and nuclear compartments of islet cells [33], or between mouse and rat β-cells.

Figure 6. Effects of glucose and ZnCl2 during culture on subsequent glucose-stimulated insulin secretion and on preproinsulin mRNA expression. Islets were cultured for 3 days in the presence of G5 or G10 alone or with 100 μmol/l ZnCl2 (A,B,E,F,I), or for 1 week in the presence of G10 or G30 alone or with 100 μmol/l ZnCl2 (C,D,G,H,J). Batches of 5 islets were then pre-incubated for 45 min in G0.5 in the absence of ZnCl2 and next incubated for 1 h in the presence of G0.5, G6, G10 or G20. The islet insulin (I,J) and DNA content were measured at the end of the incubation. Insulin secretion was normalized for differences in islet DNA content (A–D) or for differences in islet insulin content (E–H). *, P<0.05 for the acute effect of glucose vs. G0.5 and **, P<0.05 for the effect of culture in G5 or G30 vs. G10 by two-way ANOVA followed by a test of Bonferroni. I, J, P<0.05 for the effect of culture in G5 and G30 vs. G10 and **, P<0.05 for the effect of ZnCl2 during culture by two-way ANOVA followed by a test of Bonferroni. K–L, after preculture, rat islets were cultured for 1 week in G5, G10 or G30 alone (open bars) or in the presence of 100 μmol/l ZnCl2 (closed bars). Preproinsulin to cyclophilin mRNA ratios were measured by real-time PCR and expressed relative to the ratio measured in islets cultured in G10. K, the mean Ct for cyclophilin were 23.7 in G5, 24 in G5+ZnCl2, 24.1 in G10 and 24.4 in G10+ZnCl2, L, the mean Ct for cyclophilin were 23.7 in G10, 23.9 in G10+ZnCl2, 23.8 in G30 and 24 in G30+ZnCl2. Results are means ± SEM for 3 or 4 experiments. *, P<0.05 for the effect of culture in G5 vs. G10 by two-way ANOVA followed by a test of Bonferroni. doi:10.1371/journal.pone.0046831.g006
It has previously been shown that Zn\(^{2+}\) supplementation reduces early graft failure in diabetic rats transplanted with syngeneic islets [21], protects mice from diabetes induced by multiple low doses of streptozotocin [34] and ameliorates glucose tolerance in type 1 and type 2 diabetic patients [19,35], but the underlying mechanisms are not clear [20]. Also in \textit{vitro}, addition of ZnSO\(_4\) to the culture medium partially protected islet cells against the toxic effect of streptozotocin [36]. In the present study, addition of 50–100 \(\mu\)mol/l ZnCl\(_2\) to the culture medium significantly reduced mitochondrial thiol oxidation and \(\beta\)-cell apoptosis triggered by prolonged exposure to low or high \(\beta\)-intermediate glucose concentrations. These effects were unlikely due to the negligible increase in chloride anions but rather resulted from the provision of Zn\(^{2+}\), a trace element surprisingly absent from standard RPMI medium. In another study in which medium contained 5 mg/ml BSA as in ours, \(\sim 75\%\) of Zn\(^{2+}\) was bound to BSA [37]. Thus, the antiapoptotic effects of Zn\(^{2+}\) on whole islets were observed at a concentration similar to that measured in rodent plasma [37] but lower than those reported to exert proapoptotic effects in \(\beta\)-cells or other cell types [38–41]. Differences in Zn\(^{2+}\) binding or the addition of Zn\(^{2+}\) through FBS may explain the need to reduce ZnCl\(_2\) concentration at 50 \(\mu\)mol/l to avoid apoptosis of islet cell clusters. Although it has been shown that addition of 90 \(\mu\)mol/l ZnSO\(_4\) does not increase the intracellular zinc content of INS-1E cells [41], a recent study using a new zinc-sensitive fluorescent protein in mouse islet cell clusters has recently demonstrated that addition of 50 \(\mu\)mol/l ZnCl\(_2\) approximately doubled their [Zn\(^{2+}\)], while increasing \(M_{1}\) gene expression [32]. It is therefore likely that ZnCl\(_2\) treatment induces a rise in [Zn\(^{2+}\)], in rat as in mouse islets.

The beneficial effect of ZnCl\(_2\) on mitochondrial glutathione oxidation after culture in low (and to some extent high) \(\beta\)-intermediate glucose concentrations may contribute to its antiapoptotic effect in \(\beta\)-cells. Although Zn\(^{2+}\) deficiency has been shown to increase oxidative stress in other cell types, few studies have demonstrated that Zn\(^{2+}\) decreases oxidative stress (reviewed in [42]). The mechanism could involve the increase in metallothionein expression, a group of proteins that play a role in Zn\(^{2+}\) distribution to the lipids and in protection of the proteins damaged by oxidative stress [15]. \(M_{1}\)s are also known to be ROS scavengers with a capacity to capture hydroxyl radicals \(\sim 50\) times greater than that of glutathione [43]. The mechanism could also involve the role of Zn\(^{2+}\) in Cu/Zn superoxide dismutase (SOD1) activity, an enzyme which removes the superoxide radical, although it has been shown that Zn\(^{2+}\) deficiency does not decrease SOD1 activity [44,45]. In addition, other pathways, such as the inhibition of caspase 3, 6 and 9 could play a role in the antiapoptotic effect of Zn\(^{2+}\) (reviewed in [42]).

Despite the significant improvement in \(\beta\)-cell survival, Zn\(^{2+}\) did not have any protective effect on the alterations of GSIS induced by prolonged culture in extreme glucose concentrations, except for a significant increase in the insulin content of high-glucose-cultured islets. Actually, addition of ZnCl\(_2\) during culture even decreased subsequent insulin secretion in the presence of 10 mmol/l glucose, an effect that was more pronounced when insulin secretion was expressed relative to the islet insulin content.

These effects, including the increase in insulin content after culture in 30 mmol/l glucose, could result from the inhibition of Ca\(^{2+}\) influx and GSIS by Zn\(^{2+}\) in a slowly reversible manner [37,46,47]. Alternatively, we cannot exclude that the inhibition of insulin secretion by ZnCl\(_2\) in G10 indirectly resulted from an inhibition of glucagon secretion by \(\alpha\)-cells (reviewed in [48]), hence of [cAMP] in \(\beta\)-cells. However, a similar lack of improvement of GSIS was observed when islet cell apoptosis triggered by prolonged culture in 5 instead of 10 mmol/l glucose was inhibited by 50–70% with the SOD and catalase-mimetic manganese (III)tetrakis (4-benzoic acid)porphyrin [MnTBP], [49]. Two hypotheses may explain these results. Either \(\beta\)-cell function is more sensitive than survival to the remaining level of oxidative stress present in the presence of Zn\(^{2+}\) or MnTBP, or the loss of GSIS under these culture conditions is unrelated to mitochondrial oxidative stress. Thus, although an increase in \(\beta\)-cell mass with no alteration in secretory function may contribute to the beneficial effect of Zn\(^{2+}\) supplementation on glucose tolerance in diabetes, our results emphasize the importance of testing GSIS in addition to cell survival when testing potential treatments of stressed \(\beta\)-cells.

In conclusion, culture of rat pancreatic islets in either low or high \(\beta\)-intermediate glucose concentrations triggers early mitochondrial oxidative stress with \(M_{1}/\alpha/2a\) mRNA expression and late \(\beta\)-cell apoptosis with loss of GSIS. ZnCl\(_2\) reduces mitochondrial oxidative stress and rat \(\beta\)-cell apoptosis under these culture conditions without improving GSIS.

Supporting Information

Table S1 Sequences of oligonucleotide primers and PCR conditions. The specificity of sense and anti-sense primers was checked by BLAST search. The thermal cycle profile consisted of a 3 min step at 95°C to release DNA polymerase activity followed by 40 cycles of amplification (15 sec denaturation step at 95°C, 45-60-90 sec annealing step at 60–62°C, and eventual 15–30 sec extension step at 80-82-84°C). Under these conditions, PCR efficiencies were \(\sim 0.95\) to 1.0. The melting temperature \(T_{m}\) of the amplicons was systemically determined at the end of the PCR to check their specificity. Their size corresponded to that expected from published sequences, as determined by agarose gel electrophoresis. *, Islet sample cDNA input in 25 \(\mu\)l reactions (ng total RNA equivalent).

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Author Contributions

Conceived and designed the experiments: JD LPR JCJ. Performed the experiments: JD LPR AFC JCJ. Analyzed the data: JD LPR AFC JCJ. Wrote the paper: JD JCJ.

References


